

Review

Winding through the WNT pathway during cellular development and demise

F. Li¹, Z.Z. Chong¹ and K. Maiese^{1,2}

¹Division of Cellular and Molecular Cerebral Ischemia, Wayne State University School of Medicine, Detroit, Michigan and

²Departments of Neurology and Anatomy and Cell Biology, Center for Molecular Medicine and Genetics, Institute of Environmental Health Sciences, Wayne State University School of Medicine, Detroit, Michigan, USA

Summary. In slightly over a period of twenty years, our comprehension of the cellular and molecular mechanisms that govern the Wnt signaling pathway continue to unfold. The Wnt proteins were initially implicated in viral carcinogenesis experiments associated with mammary tumors, but since this period investigations focusing on the Wnt pathways and their transmembrane receptors termed Frizzled have been advanced to demonstrate the critical nature of Wnt for the development of a variety of cell populations as well as the potential of the Wnt pathway to avert apoptotic injury. In particular, Wnt signaling plays a significant role in both the cardiovascular and nervous systems during embryonic cell patterning, proliferation, differentiation, and orientation. Furthermore, modulation of Wnt signaling under specific cellular influences can either promote or prevent the early and late stages of apoptotic cellular injury in neurons, endothelial cells, vascular smooth muscle cells, and cardiomyocytes. A number of downstream signal transduction pathways can mediate the biological response of the Wnt proteins that include Dishevelled, β -catenin, intracellular calcium, protein kinase C, Akt, and glycogen synthase kinase-3 β . Interestingly, these cellular cascades of the Wnt-Frizzled pathways can participate in several neurodegenerative, vascular, and cardiac disorders and may be closely integrated with the function of trophic factors. Identification of the critical elements that modulate the Wnt-Frizzled signaling pathway should continue to unlock the potential of Wnt pathway for the development of new therapeutic options against neurodegenerative and vascular diseases.

Key words: Akt, Alzheimer's, β -catenin, Cardiomyocytes, Dishevelled, Endothelial cells, Erythropoietin, Frizzled, GSK-3 β , Myocardial infarction, Neurons, Retinal disease, Stem cells, VEG

Introduction

Wnt proteins, derived from *Drosophila Wingless* (Wg) and the mouse Int-1 genes, represent a large family of secreted cysteine-rich glycosylated proteins. This novel family of proteins are intimately involved in cellular signaling pathways that play a role in a variety of processes that involve embryonic cell patterning, proliferation, differentiation, orientation, adhesion, survival, and apoptosis (Nusse and Varmus, 1982; Melkonyan et al., 1997; Wodarz and Nusse, 1998; Smalley and Dale, 1999; Patapoutian and Reichardt, 2000; Chong and Maiese, 2004; Nelson and Nusse, 2004). Until recently, nineteen of the twenty-four Wnt genes that express Wnt proteins have been identified in the human. In addition, more than eighty target genes of Wnt signaling pathways also have been demonstrated in human, mouse, *Drosophila*, *Xenopus*, and Zebrafish. This representation encompasses several cellular populations, such as neurons, cardiomyocytes, endothelial cells, cancer cells, and pre-adipocytes.

Wnt binds to Frizzled transmembrane receptors on the cell surface to activate downstream signaling events. These involve at least two intracellular signaling pathways with two that are considered of particular importance. One pathway controls target gene transcription through β -catenin, generally referred to as the canonical pathway that involves Wnt1, Wnt3a, and Wnt8 and functions through β -catenin-dependent pathways. Another pathway pertains to intracellular calcium (Ca^{2+}) release which is termed the non-canonical or Wnt/ Ca^{2+} pathway consisting primarily of Wnt-4, Wnt-5a, and Wnt-11 that functions through non- β -catenin-dependent pathways, such as the planar cell polarity (PCP) pathway (Nusse, 1999; Salinas, 1999; Patapoutian and Reichardt, 2000; Tada and Smith, 2000) and the Wnt- Ca^{2+} -dependent pathways (Slusarski et al., 1997; Nusse, 1999; Salinas, 1999; Kuhl et al., 2000; Patapoutian and Reichardt, 2000; Katoh, 2002). Recent work has illustrated that eleven members of the Frizzled transmembrane receptors have been identified in the

human and mouse genomes that encode for seven-pass transmembrane domain-containing serpentine receptor proteins (Nelson and Nusse, 2004). If one examines the Wnt-Frizzled signaling transduction pathway, it becomes evident that these pathways play critical roles during embryonic, non-vertebrate and vertebrate development as well as tumorigenesis (Dale, 1998; Kawakami et al., 2001; Lustig and Behrens, 2003).

Multiple studies have shown the importance of Wnt-Frizzled transduction pathway in controlling the pattern of the body axis as well as the development and maturation of the central nervous system (Augustine et al., 1993; Ikeya et al., 1997; Salinas, 1999; Wheeler and Hoppler, 1999; Melton et al., 2004), cardiovascular system (Park et al., 1996; Wheeler and Hoppler, 1999; van Gijn et al., 2001), and the limbs (Kengaku et al., 1997). During embryological development, alternations of the Wnt-Frizzled pathway can lead to abnormal morphogenesis in animal models (Stark et al., 1994; Ikeya et al., 1997; Liu et al., 1999) and congenital defects in humans (Jordan et al., 2001; Rodova et al., 2002; Niemann et al., 2004). In mature tissues, the Wnt-Frizzled pathway is involved in the self-renewal of stem cells and may be responsible for the maintenance of many normal tissues (Ross et al., 2000; Reya et al., 2003; Willert et al., 2003; He et al., 2004). Studies have revealed that dysfunction of the Wnt-Frizzled pathway can lead to neurodegenerative disorders, such as Alzheimer's disease (Anderton, 1999; Grilli et al., 2003; Maiese and Chong, 2004; Chong et al., 2005d) and heart failure (Schumann et al., 2000; Haq et al., 2001; van Gijn et al., 2002). This paper will focus upon the Wnt protein and its receptor, the Wnt-Frizzled signaling pathway and components, and the significant role of the Wnt-Frizzled signaling pathway during cellular survival and injury in a broad array of cells that involve neuronal, endothelial, and cardiac cell populations.

The structure and function of Wnt proteins and their receptors

Though they can show varying degrees of sequence identity, the molecular structural characteristics that all

Wnt proteins share involve the 39-46kDa lipid-modified secreted glycoproteins containing 350-400 amino acids, with a highly conserved pattern of 23-24 cysteine residues and several asparagines-linked glycosylation sites (Nusse and Varmus, 1992; Miller, 2002). Yet, some of Wnt proteins have an additional domain, such as the *Drosophila Wg* that contains an 85-amino acid domain near the center of protein (Nusse and Varmus, 1992).

Several members of Wnt proteins have been identified to control proliferation, differentiation, and death of various cells. The cell populations can include stem cells as well as the development of various tissues that in the nervous and cardiovascular systems (Table 1). Early studies have demonstrated that ectopic expression of specific Wnt genes in *Xenopus* embryos can result in distinct phenotypes. In the C57MG mouse, transient expression of Wnt1, Wnt2 and Wnt3a in mammary epithelial cells can cause morphological transformation while the other Wnt proteins have little effect on cell morphology (Wong et al., 1994). In addition, in *Xenopus* embryos, the injection of Wnt1, Wnt3a and Wnt8 into the ventral blastomeres of four-cell embryos can lead to duplication of the body axis, but the overexpression of Wnt4, Wnt5a and Wnt11 genes can interfere with morphogenetic movement without inducing axis duplication (Smith and Harland, 1991; Sokol et al., 1991; Christian et al., 1992; Moon et al., 1993; Wolda et al., 1993).

As a result, Wnt proteins are generally divided into two functional classes based on their ability to induce a secondary body axis in *Xenopus* embryos and to activate certain signaling cascades that consist of the Wnt1 class and the Wnt5a class. The members of the Wnt1 class are inducers of a secondary body axis in *Xenopus* and include Wnt1, Wnt2, Wnt3, Wnt3a, Wnt8 and Wnt8a. Wnt proteins of this class facilitate activation of the Frizzled transmembrane receptor and the co-receptor lipoprotein related protein 5 and 6 (LRP-5/6). Ultimately, this leads to the activation of the typical canonical Wnt/ β -catenin pathway. The Wnt5a class cannot induce secondary axis formation in *Xenopus* and includes the Wnt proteins of Wnt4, Wnt5a, Wnt5b, Wnt6, Wnt7a and Wnt11. These Wnt proteins bind the

Table 1. Neuronal and cardiac expression of the Wnt and the Wnt receptor with biological function.

ORGAN	CELLULAR EXPRESSION		BIOLOGICAL RESPONSE
	Wnt	Wnt Receptor	
Brain	Neurons	Neurons	Cell regulation and cytoprotection
	Astrocytes	Astrocytes	Cell regulation and cytoprotection
	Progenitor stem cells	Progenitor stem cells	Cellular development and maturation
Vascular System	Endothelial cells	Endothelial cells	Angiogenesis (cell proliferation, differentiation, migration)
	Progenitor vascular stem cells	Progenitor vascular stem cells	Angiogenesis and cardiomyogenesis
	Vascular smooth muscle cells	Vascular smooth muscle cells	Angiogenesis, vascular remodeling, and cytoprotection
Heart	Progenitor cardiac stem cells	Progenitor cardiac stem cells	Cardiomyogenesis and cardiac conduction cell development
	Endocardial cells	Endocardial cells	Endocardial cushion formation
	Cardiomyocytes	Cardiomyocytes	Cardiac remodeling and cytoprotection

Wnt during development and demise

Frizzled transmembrane receptor to activate heterotrimeric G proteins and increase intracellular calcium levels. Alternatively, they can induce Rho-dependent changes in the actin cytoskeleton. Several recent studies also have shown that the different subsets of Wnt proteins can contribute to distinct physiological changes through triggering various intracellular pathways (Heisenberg et al., 2000; Tada and Smith, 2000; Winklbauer et al., 2001; Hsieh, 2004).

The main receptors of the Wnt proteins consist of at least 10 family members termed the Frizzled proteins after the first member, *Drosophila* tissue polarity gene I (Vinson et al., 1989; Adler et al., 1990). All members of the *Frizzled* protein family share the following characteristics: a N-terminal signal peptide, an extracellular domain that contains a 120-amino acids, a cysteine-rich domain followed by a hydrophilic linker region that shows little sequence similarity among family members, a highly conserved seven-transmembrane domain separated by short extracellular and cytoplasmic loops, and a cytoplasmic domain of variable size and little sequence homology among family members (Vinson et al., 1989; Adler et al., 1990; Wang et al., 1996; Wodarz and Nusse, 1998; Hsieh, 2004). Some Wnt proteins, such as Wnt8, can directly bind with the full-length Frizzled receptor protein. A single Wnt protein also can bind to a combination of Frizzled receptor proteins, including homologous members from a different species (Bhanot et al., 1996; Hsieh et al., 1999; Hsieh, 2004). The Wnt proteins bind to the activity sites of Frizzled receptor proteins that are relevant to either the canonical and non-canonical Wnt-Frizzled signaling pathways leading to specific biological functions. Interestingly, the Frizzled cysteine-rich domain also exists in several other proteins that include the soluble secreted Frizzled-related proteins (Finch et al., 1997; Leyns et al., 1997; Mayr et al., 1997; Melkonyan et al., 1997; Rattner et al., 1997; Wang et al., 1997a,b; Ellies et al., 2000; Jones et al., 2000), some receptors of tyrosine kinases (Jennings et al., 1993; Wilson et al., 1993; Glass et al., 1996), carboxypeptidase Z (Song and Fricker, 1997), the membrane-bound serine protease Corin (Yan et al., 1999), and an isoform of collagen (Rehn and Pihlajaniemi, 1995). These proteins appear to function as important regulators during Wnt-Frizzled signaling. For example, the soluble secreted Frizzled-related proteins have been found to function as antagonists of the Wnt-Frizzled signaling pathway when ectopically expressed in *Xenopus* embryos (Finch et al., 1997; Leyns et al., 1997; Wang et al., 1997a,b; Xu et al., 1998; Ellies et al., 2000; Jones et al., 2000). In addition, the carboxypeptidase Z has been identified to modulate Wnt 4a activity through its cysteine-rich domain (Hsieh, 2004).

In addition to the Frizzled protein receptors, other obligate co-receptors also are necessary for canonical Wnt-Frizzled signaling pathway. An additional single-pass transmembrane protein named as LRP-5/6 from the low-density-lipoprotein receptor family is required for this process (Pinson et al., 2000; Tamai et al., 2000;

Wehrli et al., 2000). In the canonical Wnt-Frizzled signaling pathway, Wnt binds to both the Frizzled transmembrane receptor and the co-receptor LRP-5/6 (Wehrli et al., 2000) resulting in the inhibition of the downstream component glycogen synthase kinase-3 β (GSK-3 β) (Ikeda et al., 1998; Papkoff and Aikawa, 1998). More recent studies also have suggested that the Wnt signaling can be transmitted through the binding of extracellular domain of LRP-5/6 to Axin, a key component in the GSK-3 β complex, indicating that the LRP-5/6 receptor is an important part of the Wnt-Frizzled signaling pathway (Mao et al., 2001; Tolwinski et al., 2003).

Another newly identified co-receptor for the canonical and non-canonical Wnt-Frizzled signaling pathway is Ryk that belongs to one of divergent members of the receptor tyrosine kinase family. Ryk not only can form a complex with Frizzled proteins such as the co-receptor LRP-5/6 resulting in activation of the canonical Wnt-Frizzled signaling pathway, but also can regulate the non-canonical Wnt-Frizzled signaling pathway through Frizzled-independent pathways (Cheyette, 2004; Bejsovec, 2005). At this time, a single Ryk gene in mammals, one Ryk gene in the nematode *C. elegans* (Inoue et al., 2004; Deshpande et al., 2005), and three Ryk genes in *Drosophila* (Yoshikawa et al., 2003) have been identified in an atypical receptor tyrosine kinase family. The molecular structure of all Ryk genes is characterized by an extracellular domain with homology to Wnt inhibitory factor-1 (WIF1), a single transmembrane-spanning sequence to which Wnt proteins bind (Schneider et al., 1999; Patthy, 2000). In addition, a conserved intracellular PDZ-binding motif exists which links Ryk to downstream molecules of Wnt-Frizzled signaling pathway, such as Dishevelled (Cheyette, 2004; Lu et al., 2004; Bejsovec, 2005).

Wnt proteins can bind to the extracellular domain of the Ryk receptor through the intracellular PDZ-binding domain in the Ryk receptor to result in regulating cell proliferation, differentiation, migration, polarity, survival, and death through either the canonical or the non-canonical Wnt-Frizzled signaling pathway. As an example, mammalian Ryk can function as a co-receptor with Frizzled to bind to Wnt1 and Wnt3a through its WIF domain and interact with Dishevelled via its PDZ domain resulting in the stimulation of neurite outgrowth (Lu et al., 2004). Furthermore, in *C. elegans*, Wnt proteins can employ Frizzled encoded by *lin-17* and Ryk encoded by *lin-18* to regulate cell fate during *C. elegans* vulval development (Inoue et al., 2004; Deshpande et al., 2005). In the *Drosophila* embryonic nervous system, Wnt5 specifically binds to the Derailed receptor that is a member of the Ryk receptor subfamily to mediate axonal guidance (Yoshikawa et al., 2003).

The Wnt-Frizzled Signaling Pathway

The Wnt-Frizzled signaling pathway includes the canonical Wnt signaling pathway and the non-canonical Wnt signaling pathway, the latter consisting of the

Wnt/PCP pathway and the Wnt/ Ca²⁺ pathway. Upon binding to either the Frizzled receptor or a receptor complex consisting of Frizzled and LRP5/6, Wnt protein can activate one of three different signaling cascades. These cascades include the canonical Wnt signaling pathway (Rattner et al., 1997; Nusse, 1999), the Wnt/PCP pathway (Rattner et al., 1997; Nusse, 1999; Veeman et al., 2003), or the Wnt/ Ca²⁺ pathway (Slusarski et al., 1997; Nusse, 1999; Salinas, 1999; Kuhl et al., 2000; Patapoutian and Reichardt, 2000; Katoh, 2002). Each of pathways, although distinct, appears to be transduced initially through Dishevelled, a cytoplasmic multifunctional phosphoprotein (Axelrod et al., 1998; Boutros et al., 1998; Boutros and Mlodzik, 1999). Yet, the ultimate response to Wnt-Frizzled interaction will most probably depend upon cellular context at that time (Ilyas, 2005). In mammals, the Dishevelled protein family members contains Dishevelled-1, Dishevelled-2, Dishevelled-3 in all organs. These family members have three highly conserved domains that include an N-terminal DIX domain named for Dishevelled and Axin, a central PDZ domain termed for Postsynaptic density-95, Discs-large and Zonula occludens-1, and a C-terminal DEP that is named for Dishevelled, Egl-10 and Pleckstrin (Wharton, 2003; Habas and Dawid, 2005). At the level of Dishevelled, the Wnt signaling pathway can be separated along one of three different cascades that are dependent upon the three highly conserved domains of Dishevelled. As a result, Dishevelled is a key transducer of the Wnt signal that acts at the plasma membrane or in the cytoplasm in all three Wnt-Frizzled signaling pathways. However, new work has suggested that Dishevelled also acts within the nucleus and nuclear location of Dishevelled is essential for its function in the Wnt-Frizzled signaling pathway (Itoh et al., 2005; Weitzman, 2005).

The canonical Wnt signaling pathway

The canonical Wnt signaling pathway is considered to be a typical Wnt-Frizzled signaling pathway. It is also referred to as the Wnt/ β -catenin-dependent signaling pathway or the Wnt/ β -catenin pathway since it can regulate β -catenin protein levels to control the activation of Wnt-responsive target genes. The majority of Wnt proteins activate gene transcription through this signaling pathway controlled by β -catenin. In general, all Wnt signaling pathways are initiated by interaction of Wnt proteins with Frizzled receptors, but in this pathway, the Wnt signaling pathway will only be activated if the binding of the Wnt protein to the Frizzled transmembrane receptor takes place in the presence of the co-receptor LRP-5/6 (Pinson et al., 2000; Wehrli et al., 2000; Mao et al., 2001) resulting in the formation of a Wnt-Frizzled-LRP5/6 trimolecular complex. Once Wnt protein binds to the Frizzled transmembrane receptor and the co-receptor LRP-5/6, this is followed by recruitment of Dishevelled. Dishevelled is

phosphorylated by casein kinase I ϵ to form a complex with Frat1 and inhibit GSK-3 β activity (Ikeda et al., 1998; Papkoff and Aikawa, 1998; Lee et al., 1999; Kishida et al., 2001; Lee et al., 2001). In addition, the formation of the Wnt-Frizzled-LRP5/6 complex also promotes the LRP5/6-mediated degradation of Axin (Mao et al., 2001).

As a result, the combined inhibition of GSK-3 β activity with the degradation of Axin blocks the formation of the protein complex consisting of GSK-3 β , Axin, and adenomatous polyposis coli (APC) tumor suppressor protein. Yet, during the absence of Wnt signaling, β -catenin is associated with the protein complex of GSK-3 β , Axin and APC tumor suppressor protein. β -catenin is phosphorylated by activation of GSK-3 β leading to its ubiquitination and subsequent degradation by proteosomes (Aberle et al., 1997; Hart et al., 1999; Latres et al., 1999; Winston et al., 1999; Patapoutian and Reichardt, 2000). As a result, β -catenin cannot translocate into the nucleus and physically bind to DNA in order to activate the transcription of its target genes. In the absence of β -catenin in the nucleus, the T cell factor (Tcf) and lymphocyte enhancer factor (Lef) (Tcf/Lef) family members are associated with transcriptional inhibitors, such as Groucho (Cavallo et al., 1998; Roose et al., 1998).

However, without the formation of the protein complex of GSK-3 β , Axin and APC tumor suppressor protein, phosphorylation of β -catenin with its subsequent degradation does not occur and the accumulation of free β -catenin results for translocation to the nucleus (Cavallo et al., 1998; Ikeda et al., 1998; Roose et al., 1998; Akiyama, 2000). Once positioned in the nucleus, the free β -catenin acts as a transcription factor and activates Tcf and Lef by forming nuclear complexes with members of the Tcf/Lef transcription factor family (Ishitani et al., 2003). This leads to the transcription and expression of a variety of Wnt-responsive target genes such as c-Myc (He et al., 1998), cyclin D1 (Nusse, 1999; Shtutman et al., 1999; Tetsu and McCormick, 1999), and Axin 2 (Jho et al., 2002; Lustig et al., 2002). In addition, the complexes of Tcf/Lef and β -catenin may cooperate with factors activated by other signaling pathways to alter cellular remodeling processes. Therefore, the Wnt/ β -catenin pathway affects a broad array of cellular functions and regulates cell fate, proliferation, differentiation, adhesion, and survival through increased β -catenin levels with gene modulation via Tcf/Lef transcription factors (Behrens et al., 1996).

The canonical Wnt signaling pathway also is activated by several other cellular mechanisms. The shifting of proteins from the cadherin-bound pool to the cytoplasmic pool can increase the amount of available free β -catenin for the activation of target genes. Several receptor tyrosine kinases can phosphorylate tyrosine residues of the β -catenin and cadherin-catenin complex to allow β -catenin to become dissociated from the complex and increase the amount of β -catenin in the cytoplasm for subsequent translocation to the nucleus

(Ilyas, 2005). Furthermore, surface receptors, such as epidermal growth factor receptor, c-ROn and cErbB2, can then stimulate the canonical Wnt signaling pathway (Bonvini et al., 2001; Danilkovitch-Miagkova et al., 2001; Graham and Asthagiri, 2004). In addition, the insulin-like growth factor causes tyrosine phosphorylation and β -catenin stabilization (Playford et al., 2000). Integrin-linked kinase also can activate the canonical Wnt signaling pathway through the inhibition of GSK-3 β and cAMP-responsive element-binding protein-dependent pathway (D'Amico et al., 2000).

The Non-Canonical Wnt signaling pathway

The non-canonical Wnt signaling pathway, also termed the atypical Wnt-Frizzled signaling pathway, has two intracellular signaling cascades that consist of the Wnt/ Ca²⁺ pathway and the Wnt/PCP pathway. In the Wnt/ Ca²⁺ pathway, Wnt protein binds to Frizzled transmembrane receptors on the cell surface resulting in several cellular processes that involve stimulation of heterotrimeric G proteins, increased intracellular Ca²⁺ release, decreased cyclic guanosine monophosphate (cGMP) levels, and activation of the two kinases Ca²⁺-calmodulin-dependent protein kinase II (CamKII) or calcineurin (CaCN) and protein kinase C (PKC). These processes can stimulate nuclear factor (NF)-AT and other transcription factors (Veeman et al., 2003; Wang and Malbon, 2003; Kuhl, 2004). Thus, the Wnt/Ca²⁺ pathway is most likely a G-protein dependent signaling pathway (Kuhl et al., 2000; Kuhl, 2004; Wang and Malbon, 2004). In the Wnt/PCP pathway, Wnt proteins bind to Frizzled transmembrane receptors on the cell surface followed by activating Rho/Rac small GTPase (Habas et al., 2003) and Jun N-terminal kinase (JNK) (Moriguchi et al., 1999) to assist in the subsequent regulation of cytoskeletal organization and gene expression (Moulin and Widmann, 2004).

It is important to note that several of the downstream proteins identified in the Wnt-Frizzled signaling pathway can independently function with proteins from other cellular systems. For example, Dishevelled is able to directly regulate JNK activity (Li et al., 1999) in addition to GSK-3 β (Boutros et al., 1998). Free β -catenin also can form a complex with α -catenin and members of cadherins family to function as a structural adaptor protein linking cadherins to the actin cytoskeleton in cell-cell adhesion processes (Jamora and Fuchs, 2002; Nelson and Nusse, 2004). Association with cadherins can effectively sequester β -catenin from the cytoplasmic pool that is responsive to Wnt-Frizzled signaling (Fagotto et al., 1996; Sadot et al., 1998; Simcha et al., 1998). Therefore, the modulation of cadherin expression and function can indirectly regulate the Wnt-Frizzled signaling pathway through β -catenin. Because both cadherins and Wnt proteins have been demonstrated to regulate aspects of synapse formation, the interaction between these proteins also may play a critical role in the development of nervous system (Hall

et al., 2000; Patapoutian and Reichardt, 2000; Tanaka et al., 2000).

Wnt signaling and cellular development in neuronal and vascular systems

The Wnt-Frizzled signaling pathway plays an important role in the biology of the development of the nervous system. In particular, the Wnt-Frizzled signaling pathway is involved in the development of the neural plate with neuronal progenitor cells and with the subsequent anterior-posterior extension of the neural tube. Ultimately, the Wnt-Frizzled signaling pathway leads to the development of the brain, spinal cord, and the extension of numerous sub-populations of sensory and motor neurons.

Several studies have demonstrated that the Wnt-Frizzled signaling pathway can regulate a number of patterning events during development of nervous system. In *Xenopus* embryos, the Wnt-Frizzled signaling pathway has been shown to activate neuronal development through inhibition of the expression of bone morphogenetic protein (BMP) 4 (Baker et al., 1999). Yet, the inhibition of BMP alone is insufficient to account for neural crest induction in *Xenopus*. Other BMPs also play a role during the development of nervous system. BMP 5 is involved in dorsal specification, which may play an important role in dorsal-ventral patterning of the developing brain which is ultimately under control of the Wnt-Frizzled signaling pathway (Golden et al., 1999; Ellies et al., 2000). Other work has revealed that BMP and a Wnt-BMP signaling loop controls cell proliferation, migration, and axonal guidance of neurons in the developing nervous system (Yeo and Gautier, 2004; Chizhikov and Millen, 2005). It also appears that blockade of Wnt8 function, as shown by overexpression of a dominant negative Wnt8, can inhibit the expression of neural crest markers, suggesting that the Wnt-Frizzled signaling pathway also is necessary for neural crest induction (LaBonne and Bronner-Fraser, 1998; Lewis et al., 2004). Other studies that employ gain-and loss-of-function for Wnt signaling have demonstrated that the Wnt-Frizzled signaling pathway plays a critical role either in neural crest induction or in the specification of the neural crest competence territory (Bastidas et al., 2004).

The canonical Wnt -Frizzled signaling pathway appears to be required for anterior neural patterning in studies with *Xenopus* embryos (Kiecker and Niehrs, 2001). *Xidax*, an inhibitor of the canonical Wnt -Frizzled signaling pathway, can reduce the expression of anterior neural markers, indicating that the canonical Wnt -Frizzled signaling pathway is crucial for the anterior neural development in *Xenopus* (Michiue et al., 2004). Interestingly, the co-expression of Wnt1 and Wnt3a may be necessary for the development of the dorsal neural tube since loss of these two Wnt proteins results in fewer dorsal lateral neural precursors, suggesting that the Wnt-Frizzled signaling pathway plays a vital role in

regulating dorsal neural patterning (Ikeya et al., 1997; Muroyama et al., 2002; Chizhikov and Millen, 2005).

In fact, several cellular proteins in the Wnt-Frizzled signaling pathway that have been shown to be involved in the dorsal-ventral patterning of the neural tube also directly regulate patterning of the telencephalon (Grove and Tole, 1999) and also can contribute to forebrain patterning in the developing brain (Braun et al., 2003; Abu-Khalil et al., 2004). For example, Wnt 3a, Wnt 5a, and Wnt 2b contribute to the development of the cortical hem which forms the boundary between the hippocampus and choroids plexus in the embryonic cerebral cortex (Grove et al., 1998; Lee et al., 2000). Wnt genes, genes encoding Frizzled Wnt receptors, or secreted Frizzled-related proteins and Tcf/Lef-1 transcription factors, also are expressed in postnatal mouse cerebral cortex lasting into young adulthood, further indicating that the Wnt/ β -catenin signaling pathway represents a major cortical input during embryonic brain development (Shimogori et al., 2004).

The Wnt-Frizzled signaling pathway also functions as a regulator of specific precursor populations in the developing brain. Ectopic expression of Wnt1 can strongly induce overproliferation of precursor cells in the caudal midbrain during midgestation. Yet, in the mature organism, Wnt 1 exhibits a cell size promoting effect specifically on neurons, suggesting modulation of the development process during different periods of gestation by the same Wnt protein (Panhuysen et al., 2004). Additional work also demonstrates that autoregulation of canonical Wnt/ β -catenin signaling pathway can control midbrain development through the expression of transcription factor Tcf-4 isoforms that require Wnt2b, but also control Wnt2b (Kunz et al., 2004). It also is clear that Lef1/Tcf proteins regulate the generation of dentate gyrus granule cells and the development of the hippocampus (Galceran et al., 2000). Mouse embryos homozygous for a Lef1-lacZ fusion gene, which encodes a protein that not only is deficient in DNA binding, but also interferes with β -catenin-mediated transcriptional activation by other Lef1/Tcf proteins, are absent of the hippocampal structure. Recent studies further demonstrate roles for Dishevelled, Rac, and JNK signaling pathways during neuronal development. Wnt7 β and Dishevelled can activate Rac and JNK signaling pathways to promote dendritic branching growth in cultured hippocampal neurons, since application of dominant-negative Rac, administration of dominant-negative JNK, or inhibition of JNK activity can inhibit Dishevelled-mediated dendritic growth (Rosso et al., 2005). These multiple studies illustrate that the Wnt-Frizzled signaling pathway can employ β -catenin and Lef1/Tcf transcription factors to control the proliferation and differentiation of almost all cells of nervous system. The Wnt-Frizzled signaling pathway regulates the development of the nervous system through multiple cell populations that can involve neural stem cells (Bronner-Fraser, 2004; Hirabayashi et al., 2004; Lee et al., 2004;

Muroyama et al., 2004), cortical and hippocampal neurons (Patapoutian and Reichardt, 2000; Machon et al., 2003), dopaminergic neurons (Castelo-Branco et al., 2003), and sensory neurons (Bronner-Fraser, 2004; Lee et al., 2004).

In consideration of the extensive involvement of the Wnt system during development of the brain, it should come as no surprise that Wnt proteins also have a significant role in the vascular system. In work that has examined β -catenin expression in the avian mesonephros, a transitory embryonic kidney that is used in the study of vascular development and degeneration, degenerating mesonephros and glomerular capillary tufts had significantly depressed β -catenin expression. This was in contrast to viable cells with prominent β -catenin expression, suggesting that β -catenin expression was linked to remodeling of the vascular system (Nacher et al., 2005). Other studies show that targeted disruption of LRP-5, a Wnt co-receptor, results in persistent embryonic eye vascularization, further supporting a role for Wnt during postnatal vascular regression (Kato, 2002). In regards to cardiac development, it has been shown that conditional targeting of APC, a protein that can down-regulate intracellular levels of β -catenin, in the neural crest yields apoptosis of cardiac neural crest cells, resulting in cardiac anomalies at birth (Hasegawa et al., 2002). This work suggests that Wnt signaling and β -catenin is required for proper development of cardiac tissue. Several studies also provide support for the ability of the Wnt system through GSK-3 β to regulate cardiac development and hypertrophy (Hardt and Sadoshima, 2002). Additional studies demonstrate that Wnt 11 can promote cardiomyogenic differentiation of human circulating endothelial progenitor cells through activating the non-canonical PKC-dependent signaling pathway (Koyanagi et al., 2005).

Control of apoptotic pathways by Wnt signaling

Apoptosis, also termed programmed cell death, is considered to be an active component of cell death that contributes to neuronal, myocardial and vascular destruction. Apoptosis, along with the proliferation and differentiation of cells, has long been recognized to play an important role in processes such as tissue homeostasis and morphogenesis during normal vertebrate development. During the development of the nervous system, the neurotrophic apoptosis that occurs within undifferentiated and differentiated cells has served as a paradigm to illustrate the importance of apoptosis for the normal development of nervous system (Yeo and Gautier, 2004). Dysfunctions in the regulation or execution of apoptosis are implicated in a wide range of developmental abnormalities and diseases (Maiese and Chong, 2004; Mattson, 2004). Apoptosis also serves as a central pathway that can lead to a cell's demise in a variety of tissues and has recently been identified in organisms as diverse as plants (Hatsugai et al., 2004).

Apoptosis consists of membrane phosphatidylserine

Wnt during development and demise

(PS) exposure and DNA fragmentation (Maiese et al., 2004) (Fig. 1). Apoptotic injury is believed to contribute significantly to a variety of disease states that especially involve the nervous system such as cerebral ischemic disease, Alzheimer's disease, and trauma (Chong et al., 2004b; Doonan and Cotter, 2004; Ferretti, 2004; Koyama and Ikegaya, 2004; Li et al., 2004a). Outside of the nervous system, such as during cardiovascular injury, apoptosis also may be a significant precipitant of cell death (Maiese, 2001; Maiese and Chong, 2003), since ischemic injury has been shown to result in apoptosis in cardiomyocytes (Cai et al., 2003).

As an early event in the dynamics of cellular apoptosis, PS exposure may be required for embryogenesis (Bose et al., 2004). Yet, in mature tissues, membrane PS externalization can become a signal for the phagocytosis of cells (Hong et al., 2004; Chong et al., 2005a) (Fig. 1). In the nervous system, cells expressing externalized PS may be removed by microglia (Lin and Maiese, 2001; Li et al., 2004b). An additional role for membrane PS externalization in the vascular cell system is the activation of coagulation

casades (Chong et al., 2002b, 2004a). The externalization of membrane PS residues in endothelial cells (ECs) can promote the formation of a procoagulant surface (Bombeli et al., 1997) (Fig. 1).

In contrast to the early externalization of membrane PS residues, the cleavage of genomic DNA into fragments is considered to be a delayed event that occurs late during apoptosis (Dombroski et al., 2000; Maiese and Vincent, 2000; Jessel et al., 2002; Kang et al., 2003b). Several enzymes responsible for DNA degradation have been differentiated based on their ionic sensitivities to zinc (Torriglia et al., 1997) and magnesium (Sun and Cohen, 1994). Calcium, a critical independent component that can determine cell survival (Weber, 2004), also may determine endonuclease activity through calcium/magnesium - dependent endonucleases such as DNase I (Madaio et al., 1996). Other enzymes that may degrade DNA include the acidic, cation independent endonuclease (DNase II) (Torriglia et al., 1995), cyclophilins (Montague et al., 1997), and the 97 kDa magnesium-dependent endonuclease (Pandey et al., 1997). In the nervous system, three separate endonuclease activities are present that include a constitutive acidic cation-independent endonuclease, a constitutive calcium/magnesium-dependent endonuclease, and an inducible magnesium dependent endonuclease (Vincent and Maiese, 1999b). The physiologic characteristics of the magnesium dependent endonuclease, such as a pH range of 7.4-8.0, a dependence on magnesium, and a molecular weight of 95-108 kDa, are consistent with a recently described constitutive 97 kDa endonuclease in non-neuronal tissues.

Oxidative stress can lead to apoptosis in neurons, ECs, cardiomyocytes, and smooth muscle cells through multiple cellular pathways (Chong et al., 2005c). Oxidative stress, such as nitric oxide (NO) exposure results in nuclei condensation and DNA fragmentation (Vincent and Maiese, 1999b; Goldshmit et al., 2001; Chong et al., 2003b; Pugazhenthil et al., 2003). In neurons, NO exposure produces apoptotic death in hippocampal and dopaminergic neurons (Vincent and Maiese, 1999a; Witting et al., 2000; Chong et al., 2003a; Sharma and Ebadi, 2003). Injury during NO exposure also can become synergistic with hydrogen peroxide to render neurons more sensitive to oxidative injury (de la Monte et al., 2003; Wang et al., 2003). Externalization of membrane PS residues also occurs in neurons during anoxia (Chong et al., 2002b), NO exposure (Chong et al., 2003d), or during the administration of agents that induce the production of reactive oxygen species, such as 6-hydroxydopamine (Salinas et al., 2003).

The Wnt-Frizzled signaling pathway controls apoptosis in a variety of cell populations during both development and injury (Fig. 2). During development, Wnt signaling can either facilitate or prevent apoptosis depending upon the environmental stimuli present. For example, Wnt proteins can regulate apoptosis within rhombomeres 3 and 5 in the developing hindbrain and in

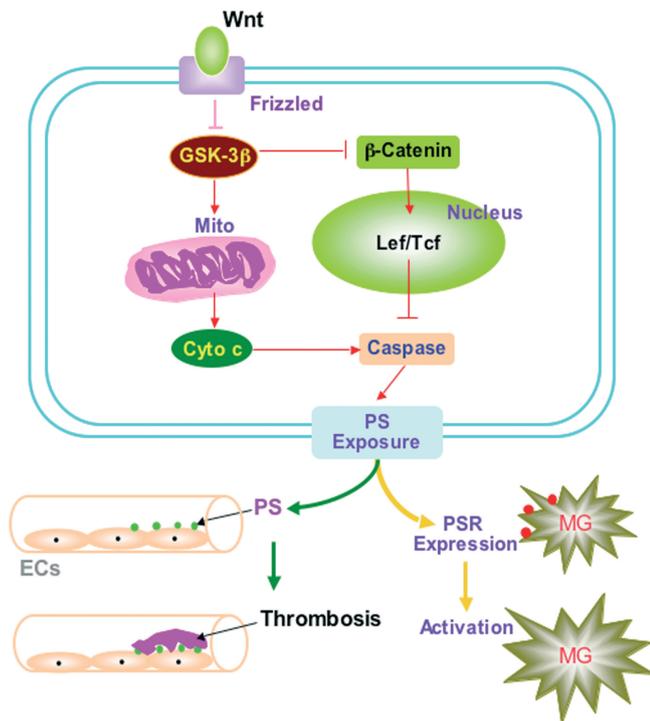


Fig. 1. Wnt maintains cellular integrity in endothelial cells and potentially fosters a novel capacity to block thrombosis and microglial activation. Wnt functions through activation of its receptor Frizzled resulting in the inhibition of glycogen synthase kinase-3β (GSK-3β), which induces mitochondrial (Mito) membrane depolarization followed by cytochrome c (Cyto c) release and prevents the activity of β-catenin. Without Wnt, GSK-3β can activate caspases and subsequently leads membrane phosphatidylserine (PS) exposure, resulting in the induction of thrombosis in vascular system and PS receptor (PSR) expression on microglia with microglial activation.

limb buds during vertebrate limb development to control development of the hindbrain and the limb (Ellies et al., 2000; Grotewold and Ruther, 2002a,b).

A number of studies in a variety of organisms have demonstrated that the Wnt-Frizzled signaling pathway can regulate apoptosis through a variety of mechanisms that include the Wnt-BMP signaling loop (Golden et al., 1999; Ellies et al., 2000), secreted Frizzled-related protein-2 (SFRP2) expression (Ellies et al., 2000; Jones et al., 2000), Wnt- β -catenin signaling (Galceran et al., 2000; Brault et al., 2001; Ahmed et al., 2002; Hari et al., 2002), c-Jun N-Terminal kinase signaling (Grotewold and Ruther, 2002b; Lisovsky et al., 2002; Yeo and Gautier, 2004), GSK-3 β -NF- κ B signaling (Bournat et al., 2000; Kozlovsky et al., 2002) and gene expression such as human Dickkopf-1 (hDkk-1) (Shou et al., 2002), nemo (Mirkovic et al., 2002), sox 10 (Honore et al., 2003) and tau (Jackson et al., 2002).

As one of the best characterized members of the Wnt family, Wnt1 signaling has been associated with the control of apoptosis during injury in some cell systems. Wnt1 prevents apoptosis through β -catenin/Tcf transcription mediated pathways (Chen et al., 2001; Rhee et al., 2002). Overexpression of exogenous Wnt1 results in the protection of cells against c-myc induced apoptosis through induction of β -catenin, cyclooxygenase-2, and Wnt1 induced secreted protein

(WISP-1) (You et al., 2002). Wnt1 signaling also can inhibit apoptosis through prevention of cytochrome c release from mitochondria and the subsequent inhibition of caspase 9 activation (Chen et al., 2001). The APC gene also appears to represent another mechanism that regulates apoptosis. The APC gene functions to cleave β -catenin leading to the down-regulation of transactivation of Tcf/Lef (Munemitsu et al., 1995). Without Tcf/Lef activity, APC is then permitted to increase the activities of caspase 3, caspase 7, and caspase 9 and lead to the cleavage of poly (ADP-ribose) polymerase (PARP) to enhance the vulnerability of cells to apoptosis (Chen et al., 2003).

SFRP2 is one of the family members of secreted Frizzled-related proteins, also known as secreted apoptosis-related protein 1 (SARP1), that functions as an antagonist of the Wnt-Frizzled signaling pathway to reduce apoptosis during development (Melkonyan et al., 1997; Rattner et al., 1997). For example, there exists a negative relationship between the expression of SFRP2 and the occurrence of apoptosis in rhombomeres 3 and 5. The overexpression of SFRP2 in the rhombencephalic neural crest can prevent the apoptosis of premigratory neural crest cells from rhombomeres 3 and 5 by inhibiting the expression of Wnt1 and BMP 4. In contrast, depleting SFRP2 function or overexpressing Wnt1 in rhombomeres results in apoptosis (Ellies et al., 2000). As an example of how the cellular environment can influence the modulation of apoptosis by the Wnt system, SFRP2 can be associated with increased apoptotic injury during degenerative disease processes such as retinitis pigmentosa that is characterized by progressive death of the photoreceptors due to apoptosis. SFRP2 has been shown to have significantly increased expression in areas of retinal degeneration, suggesting that during neurodegenerative disease blockade of Wnt signaling can lead to cell injury (Jones et al., 2000). In a similar vein, human Dkk-1 and Nemo also are Wnt antagonists similar to the SFRPs, but they can contribute to promote apoptosis by inhibiting the Wnt-Frizzled signaling pathway (Mirkovic et al., 2002; Shou et al., 2002). The results of exogenous application of Dkk-1 have identified that the apoptosis in the limb bud is induced by Dkk-1 through the prevention of the Wnt - Frizzled signaling pathway (Glinka et al., 1998; Grotewold and Ruther, 2002b).

The Wnt- β -catenin signaling pathway is a typical Wnt-Frizzled signaling pathway that regulates multiple cellular and development processes including control of cell proliferation and differentiation, cell polarity, and specification of cell fate, such as apoptosis. Yet, different from the Wnt-BMP signaling loop, the Wnt- β -catenin signaling pathway can prevent apoptosis through the regulation of β -catenin and Tcf/Lef. In β -catenin mutant embryos, the removal of β -catenin can lead to apoptotic loss of the hindbrain, the melanocyte lineage, neural crest cells, sensory neurons and dorsal root ganglia (Brault et al., 2001; Hari et al., 2002). Over-expression of exogenous Wnt-1 results in the protection of cells

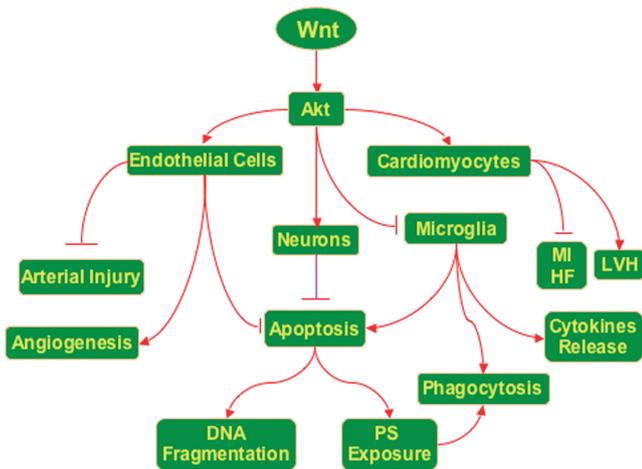


Fig. 2. The Wnt-Frizzled signaling pathway employs Akt to regulate an array of vital cellular functions that involve cellular protection, angiogenesis, cardiovascular remodeling, DNA repair, and the maintenance of membrane asymmetry in neuronal and vascular systems. The Wnt-Frizzled signaling pathway employs Akt to impact upon the survival of neurons, endothelial cells, and cardiomyocytes to prevent cellular injury and promote angiogenesis. Through the activation of Akt, microglial activity is modulated to block harmful cytokine release and prevent the phagocytosis of cells "tagged" by cellular phosphatidylserine (PS) membrane exposure. The Wnt-Frizzled signaling pathway function in cardiovascular remodeling that involves regulation of LVH (left ventricular hypertrophy) and reduction in MI (myocardial infarction) and HF (heart failure).

Wnt during development and demise

against c-Myc induced apoptosis through induction of β -catenin, cyclooxygenase-2, and Wnt-1 induced secreted protein (WISP-1) (You et al., 2002).

Additional work has revealed that the Wnt signaling pathway can decrease apoptosis and increase survival of neurons or neuronal cell lines by activating NF- κ B (Bournat et al., 2000), inhibiting GSK-3 β (Kozlovsky et al., 2002), or blocking the release of cytochrome c. In

the absence of Wnt activity, GSK-3 β phosphorylates β -catenin at serine or threonine residues of the N-terminal region to predispose degradation of β -catenin through ubiquitination (Fig. 3). GSK-3 β dependent phosphorylation of β -catenin can be promoted through phosphorylation of Axin (Yamamoto et al., 1999). In studies with chemotherapeutic agents, Wnt1 signaling also can inhibit apoptosis through prevention of cytochrome c release from mitochondria and the subsequent inhibition of caspase 9 activation (Chen et al., 2001).

The Wnt pathway also may rely upon protein kinase B (Akt) to promote cellular differentiation and survival (Chong et al., 2005b) (Fig. 3). Since Wnt can inactivate GSK-3 β and block the phosphorylation of β -catenin (Ikeda et al., 1998; Papkoff and Aikawa, 1998), this leads to the activation of β -catenin followed by transcription of its target genes for cellular protection. Akt may be necessary in pathways that involve Wnt1, since Akt inhibits the activity of GSK-3 β through phosphorylation of this protein to promote cell survival (Crowder and Freeman, 2000). Furthermore, neuronal cell differentiation that is dependent upon Wnt signaling appears to become stalled without Akt phosphorylation and the subsequent inactivation of GSK-3 β (Fukumoto et al., 2001). In addition, Wnt has been demonstrated through WISP-1 to activate the anti-apoptotic signaling pathway of Akt following genomic DNA damage (Su et al., 2002) and to block cell injury during serum withdrawal through increased Akt phosphorylation and activity (Longo et al., 2002).

Wnt and neurodegenerative disease

In line with the "anti-apoptotic" effects of Wnt signaling, absence or dysfunction in Wnt signaling can lead to neuronal injury. Wnt1 expression has been demonstrated in the brains of individuals affected by neuropsychiatric disorders (Miyaoaka et al., 1999). Furthermore, retinal degeneration during retinitis pigmentosa with the progressive loss of photoreceptors has been associated with increased secretion of Frizzled-related protein-2, a Wnt inhibitory protein, suggesting that loss of Wnt signaling may contribute to retinal neurodegeneration (Jones et al., 2000). Additional work demonstrates that a mutation in the membrane-type Frizzled-related protein gene may be involved in retinal photoreceptor degeneration (Kameya et al., 2002).

During Alzheimer's disease, neurotoxicity of A β in hippocampal neurons has been linked to increased levels of GSK-3 β and loss of β -catenin. Decreased production of Ab can occur during the enhancement of PKC activity (Savage et al., 1998) which may be controlled by the Wnt pathway (Garrido et al., 2002). The proteolytic processing of amyloid precursor protein (APP) during Alzheimer's disease also has been closely linked to the Wnt pathway through presenilin 1 (PS1) and Dishevelled. PS1 is required for the processing of APP and has been shown to down-regulate Wnt signaling and

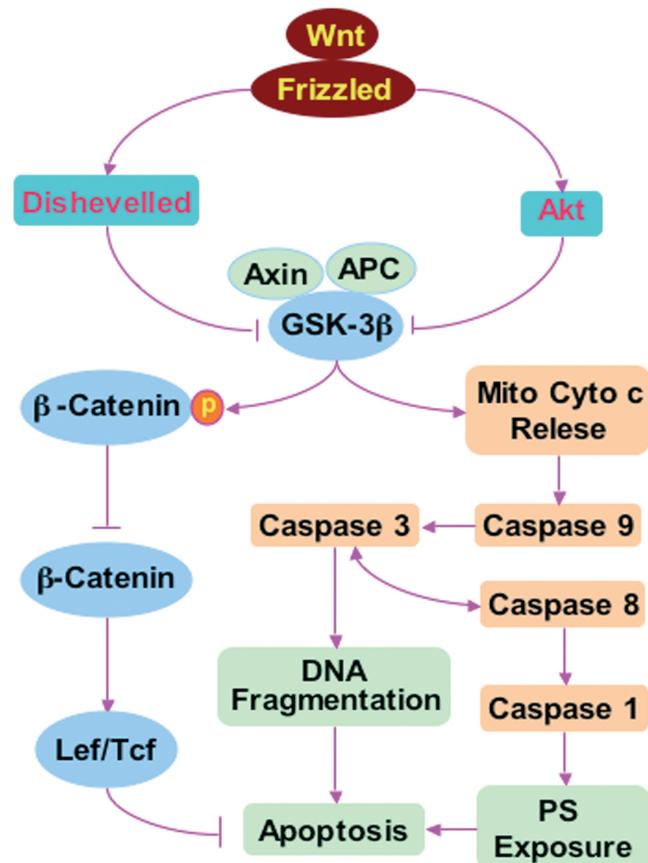


Fig. 3. The Wnt-Frizzled signaling pathway regulates PS exposure and apoptosis. The Wnt protein binds to its Frizzled receptors resulting in the activation of Dishevelled followed by the inhibition of GSK-3 β (glycogen synthase kinase), Axin and APC (adenomatous polyposis coli) tumor suppressor protein complex. The suppressed GSK-3 β , Axin and APC complex prevents phosphorylation (p) of β -catenin and leads to the accumulation of β -catenin. β -catenin enters into cellular nucleus and contributes to the formation of Lef/Tcf (lymphocyte enhancer factor/T cell factor) and β -catenin complex that may cooperate with factors activated by other signaling pathways resulting in cellular proliferation, differentiation, survival and apoptosis through the induction of target nuclear gene transcription. In addition, the binding of Wnt to Frizzled receptors also can directly activate Akt that prevents the activity of GSK-3 β and blocks Mito (mitochondrial) Cyto (cytochrome) c release. The inhibition of Cyto c release from Mito leads to the reduction of caspase activity followed by the decrease of DNA fragmentation and phosphatidylserine (PS) exposure resulting in the prevention of apoptosis.

interact with β -catenin to promote its turnover (Soriano et al., 2001). Dishevelled also can regulate the α -secretase cleavage of APP through PKC/mitogen-activated protein kinase dependent pathways, increasing soluble production of APP (sAPP) (Mudher et al., 2001). Overexpression of mouse Dishevelled-1 and -2 inhibits GSK-3 β mediated phosphorylation of tau protein and may thus prevent formation of neurofibrillary tangles during Alzheimer's disease (Wagner et al., 1997). Thus, Dishevelled may increase neuronal protection during neurodegenerative disorders through sAPP production and reduction in tau phosphorylation.

Wnt, Vascular regeneration, and vascular injury

Complimentary to the potential protective role the Wnt system may provide in neuronal populations is the ability of the Wnt pathway to modulate angiogenesis (Table 2), a process that consists of new capillary formation from pre-existing vessels into an avascular area (Chong et al., 2002a). This process involves vascular basal lamina formation, migration of ECs, and alignment of migrating cells for tubular formation

(Chong et al., 2002c; Li et al., 2004b; Sakamaki, 2004). There are at least two types of angiogenesis. Sprouting angiogenesis is characterized by the proliferation and migration of ECs into vascular sites (Risau, 1997). In contrast, non-sprouting angiogenesis or intussusceptive microvascular growth occurs by splitting the existing vasculature into transluminal pillars or transendothelial bridges (Burri and Djonov, 2002). Angiogenesis takes place in various physiological and pathophysiological conditions. It is physiologically active during embryogenesis (Risau, 1997). In the adult, it occurs during more limited periods such as during menstruation and during some pathological conditions such as wound healing, chronic inflammation, and tumor growth.

Several Wnt ligands, such as Wnt2, Wnt5a, Wnt7a, and Wnt10b, are expressed endogenously in ECs and vascular smooth muscle cells (VSMCs). More importantly, Wnt receptors that involve Fzd1, Fzd2, Fzd3, and Fzd5, as well as cysteine-rich 61 that contains Wnt-induced secreted proteins-1, 2 and 3 are also expressed in these cell populations for Wnt to exert a direct biological effect (Brigstock, 2002; Goodwin and D'Amore, 2002). It has been demonstrated that mice

Table 2. Wnt-Frizzled signaling pathway in the cardiovascular system.

CARDIOVASCULAR EVENTS	Wnt SIGNALING COMPONENTS	OUTCOME	REFERENCES
Angiogenesis	Wnt1 expression increased	HUVEC proliferation inhibited; HUVEC morphology altered	Cheng et al., 2003
	Wnt1 and β -catenin expression increased	EC and VSMC proliferation increased	Wright et al., 1999
	FrzA expression increased	Vessel density increased	Barandon et al., 2003; Dufourcq et al., 2002
	GSK-3 β inhibited and β -catenin increased	New vessel formation increased	Holthoner et al., 2002
	Fzd 4 increased; CAMKII and PKC activated increased	Retinal angiogenesis increased	Robitaille et al., 2002
Arterial Injury	Dishevelled expression and β -catenin	Neovascularization increased	Blankestijn et al., 2000
	CYR 61 expression increased	VSMC adhesion and chemotaxis increased; angiogenesis increased	Brigstock et al., 2002
Myocardial Infarction	β -catenin accumulation increased	VSMC survival increased and apoptosis decreased	Wang et al., 2002
	FrzA expression increased	Proliferating vascular ECs decreased	Duplaa et al., 1999
Cardiac Hypertrophy	FrzA expression increased	Infarct size, leukocyte infiltration, and apoptosis decreased; capillary density increased, and cardiac function improved	Barandon et al., 2003
	GSK-3 β expression inhibited	Cardiomyocyte apoptosis decreased	Bergmann et al., 2004
Cardiac Injury	GSK-3 β expression inhibited	Cardiomyocyte hypertrophy increased	Haq et al., 2000
	SFRP 3/4 increased and β -catenin decreased	Pro-apoptotic Fas increased; anti-apoptotic Bcl-xL decreased; cardiomyocyte apoptosis increased	Schumann et al., 2000
	FrzA expression increased	Cardiac apoptosis decreased; cardiac function improved	Barandon et al., 2003

CAMKII, calcium/calmodulin-dependent protein kinase II; CYR 61, cysteine-rich 61; EC, endothelial cell; Fzd, Frizzled; GSK-3 β , glycogen synthase kinase-3 β ; HUVEC, human umbilical vein endothelial cell; PKC, protein kinase C; SFRP, secreted Frizzled related protein; VSMC, vascular smooth muscle cell.

Wnt during development and demise

deficient in *Wnt2* and *Fzd5* display vascular abnormalities that include defective placental vasculature as well as embryonic lethal mutations (Ishikawa et al., 2001; Goodwin and D'Amore, 2002), suggesting that the Wnt pathway is essential for vessel development.

In several vascular cell populations, Wnt signaling plays a significant part in the modulation of new vessel formation. Overexpression of *Wnt1* can lead to the proliferation of cultured primary ECs, increase the free pool of β -catenin, and activate transcription through *Tcf/Lef*, suggesting that the Wnt-Frizzled signaling pathway is closely involved in the proliferation of ECs (Wright et al., 1999). Yet, *Wnt1* signaling also can have a regulatory role, such as during the proliferation of umbilical vein ECs, to block further growth through mechanisms that involve cell-cell contact (Cheng et al., 2003). Studies with mice homozygous for the deletion of the Wnt receptor ligand *Fzd5* that can synergize with *Wnt2*, *Wnt5a*, and *Wnt10b* lead to embryos that died in utero approximately 10 days post coitum as a result of defects in yolk sac angiogenesis, supporting a critical role for Wnt during embryo vascular development (Ishikawa et al., 2001). In contrast, mice heterozygotes were found to be viable, fertile and appear normal. In disease that involve failure of peripheral retinal vascularization, mutations in *Fzd4*, a gene encoding the Wnt receptor Frizzled-4, are believed to account for the vascular failure. Studies have shown that injection of only wildtype *Fzd4*, but not mutated *Fzd4*, into *Xenopus* embryos can activate *CamKII* and *PKC*, components of the Wnt/ Ca^{2+} signaling pathway, to control retinal angiogenesis (Robitaille et al., 2002), supporting that Wnt-Frizzled signaling pathway broadly controls vascular development and function in a number of organ systems. In addition, *FrzA*, as a member of secreted Frizzled-related protein, can increase migration and tube formation of ECs to result in enlarged, longer, and mature of vessels, further supporting the necessity of the Wnt-Frizzled signaling pathway during development of vasculature (Dufourcq et al., 2002).

In addition to vascular development, the Wnt pathway participates in the remodeling of vascular structure and the regulation of apoptosis during vascular injury (Table 2). Using a rat aorta balloon injury model, the Frizzled receptor (*rFrz*) genes, *rFrz1* and *rFrz2*, have been shown to be transiently down-regulated as early as one hour following balloon. Yet, *Frzb-1*, a secreted protein that acts as an antagonist of Wnt signaling, can be increased and appears to coincide with the arrest of aortic smooth muscle cell proliferation (Mao et al., 2000). Similarly, the secreted protein *FrzA* can be elevated in ECs during traumatic manipulation and subsequently block the proliferation of ECs (Duplaa et al., 1999). In regards to apoptotic vascular injury, it has been shown that within eight hours following EC shear stress, expression of β -catenin is significantly increased at the cell-cell junctions of ECs (Noria et al., 1999). In addition, transfection of a degradation-resistant β -catenin

into rat VSMCs blocked apoptosis during vascular balloon injury and was associated with cell cycle progression by activating cyclin D1. Furthermore, use of a dominant negative *Tcf-4* transgene lacking the β -catenin binding domain, *Tcf4(N31)*, abolished cytoprotection (Wang et al., 2002). The Wnt-Frizzled signaling pathway also is involved with venous injury. In a crush injury vein model, *Wnt5a* can have increased expression that may be beneficial coupled to the repression of inhibitor proteins of the Wnt system (Price et al., 2004).

Wnt and cardiac infarction

In regards to ischemic heart disease, new vessel formation through angiogenesis is considered to be critical for effective reparative processes (Table 2). The neovascularization following myocardial infarction contributes to the restoration of blood supply to the infarct area. In transgenic mice following myocardial infarction, overexpression of *FrzA* can result in the reduction of infarct size, prevention of cardiac rupture, and improvement in cardiac function following the increase of capillary density and the decrease of apoptotic cardiomyocytes (Barandon et al., 2003). Additional work also has demonstrated that β -catenin is translocated from the plasma membrane to the cytoplasm of ECs following the expression of *Dishevelled* during the phase of the neovascularization after myocardial infarction (Blankesteyn et al., 2000). These data suggest that the Wnt system and its components that involve β -catenin can prevent cardiomyocyte injury and preserve potential myocardial function.

Following myocardial infarction, the remodeling of the injured heart tissue involves cellular proliferation, migration, and neovascularization. The proliferation and migration of fibroblast-like cells into the infarcted area can contribute to the deposition of extracellular matrix to promote granulation tissue extension. Subsequently, myofibroblasts emerge that contribute to the preservation of cardiac function by preventing the dilatation of the infarcted area (Gabbiani, 1998). The neovascularization following myocardial infarction serves to restore blood supply to the infarcted area to preserve cardiac function.

During the complicated process of wound healing in the heart, abnormalities of some components of the Wnt-Frizzled signaling pathway, such as Wnt (Barandon et al., 2003), Frizzled (van Gijn et al., 2001), *Dishevelled-1* (Chen et al., 2004), *GSK-3 β* (El Jamali et al., 2004), β -catenin (Bergmann et al., 2004), and *SFRP* (Barandon et al., 2003), have been observed. For example, during myoblast proliferation and migration following myocardial infarction, elevated expression of Frizzled genes including Frizzled 1, 2, 5, 6, 7, 8, and 10 have been identified during heart remodeling. The overexpression of some Frizzled genes may be associated with reduction in infarct size and the

prevention of cardiac rupture to improve cardiac function (Barandon et al., 2003). It is important to note that different members of the Wnt family may contribute to distinct events during cardiac injury. Both Wnt8a and Wnt10b are upregulated following myocardial infarction while Wnt7b is down-regulated to undetectable levels during this period (Barandon et al., 2003).

Furthermore, the expression of another component, Dishevelled-1, of the Wnt-Frizzled signaling pathway also has been found in the myofibroblasts during cardiac infarction. Absence of Dishevelled-1 can not only induce the formation of myocardial infarction in mice, but also enhance the development of infarct rupture in the first week after myocardial infarction (Blankesteyn et al., 1999). In addition, the expression of Dishevelled-1 mRNA and cytoplasmic Dishevelled-1 protein are significantly enhanced within days in the myofibroblasts, vascular ECs, and smooth muscle cells of newly formed and pre-existing blood vessels in the region of a myocardial infarction. As a result, Dishevelled-1 may play a critical role in the wound healing process following myocardial infarction through the regulation of proliferation and migration of myofibroblasts and vascular ECs (Chen et al., 2004).

In addition to Frizzled, Dishevelled-1, and several Wnt genes, other members of the Wnt signaling cascade also appear to have relevant roles during myocardial infarction. Expression of SFRP3 and SFRP4 in failing human ventricular myocardium is increased when compared to donor hearts. This increased expression of SFRP is associated with the expression of the pro-apoptotic Fas proteins, but inversely linked to the expression of the anti-apoptotic protein Bcl-x_L (Schumann et al., 2000). Other studies illustrate that infarct rupture following myocardial infarction is correlated with decreased levels of β -catenin in cardiomyocytes (Blankesteyn et al., 1999). The lack of β -catenin in the adherence junctions of cardiomyocytes may lead to impaired structural integrity of the heart, since β -catenin may play a vital role in a structural adaptor protein linking cadherins to the actin cytoskeleton in cell-cell adhesion (Ligon et al., 2001). Yet, the role of β -catenin during cardiac infarction is not entirely clear, since cytosolic β -catenin exists in vascular ECs and smooth muscle cells that reside in the area of myocardial infarction (Barandon et al., 2003).

As an additional parameter that can lead to cardiac failure, pathological cardiac hypertrophy may be dependent upon modulation of the Wnt pathway. In volume-overloaded rabbit hearts, the cardiac hypertrophy is accompanied by suppressed mRNA expression of β -catenin (Itoh et al., 2002). The suppressed levels of β -catenin may be a result of increased GSK-3 β activity. Ectopic expression of GSK-3 β can lead to embryonic heart development (Marvin et al., 2001; Schneider and Mercola, 2001), suggesting that GSK-3 β may function as a negative regulator of cardiac

hypertrophy. Additional work has demonstrated that inhibition of GSK-3 β by Akt through phosphorylation of a serine residue at position 9 can play an important role in the development of cardiac hypertrophy (Haq et al., 2000; Hardt and Sadoshima, 2002).

Wnt and trophic factor regulation

Interestingly, the angiogenic effects of some growth factors may be closely linked to the Wnt-Frizzled signaling pathway. Vascular endothelial growth factor (VEGF) can initiate increased EC growth and stimulate new vessel formation by promoting tyrosine phosphorylation of β -catenin in ECs (Cohen et al., 1999; Roura et al., 1999). In addition, fibroblast growth factor 2 (FGF2) can inhibit GSK-3 activity, augment nuclear levels of β -catenin, and enhance Tcf/Lef-dependent transcription of a cyclin D1-luciferase construct, suggesting that the angiogenic properties of FGF2 are tightly regulated by β -catenin activation in the Wnt-Frizzled signaling pathway (Dono et al., 2002; Holnthoner et al., 2002).

Other cytokines and trophic factors also may have some dependence on the Wnt signaling pathway (Maiese et al., 2004, 2005). One potential candidate is erythropoietin (EPO). EPO is currently approved by the Food and Drug Administration for the treatment of anemia that can be the result of a variety of conditions. Yet, in a relatively short time period, the premise that EPO is required only for erythropoiesis has been advanced by studies demonstrating the existence of EPO and its receptor in other organs and tissues outside of the liver and the kidney, such as the heart and brain. As a result, EPO has been identified as a possible candidate in the formulation of therapeutic strategies for both cardiac and nervous system diseases.

EPO appears to be efficacious during cardiac ischemia and reperfusion injury (Li et al., 2004a; Maiese et al., 2005). EPO administration either prior to or during myocardial ischemia/reperfusion can decrease infarct size, resulting in enhanced cardiac function and improved left ventricular contractility (Parsa et al., 2004). EPO also can promote microvascular growth in the heart, suggesting that functional cardiac recovery with EPO may ensue also from the generation of new blood vessels (Ribatti et al., 1999). Other experimental studies have illustrated a potential direct protection of myocardial cells during EPO administration that can increase cardiac cell proliferation in neonatal rats, reduce myocardiocyte apoptosis during ischemia-reperfusion injury, and improve left ventricular function (Moon et al., 2003; Parsa et al., 2003).

In the nervous system, both cell culture and animal model work have demonstrated neuronal and vascular protection with EPO (Chong et al., 2002a,c; Genc et al., 2004). Systemic administration of EPO before or immediately after a retinal insult can protect retinal ganglion cells from apoptosis (Grimm et al., 2002) and can improve functional outcome and reduce lipid

Wnt during development and demise

peroxidation during spinal cord injury (Kaptanoglu et al., 2004). EPO also can block microglial cell activation and proliferation to prevent phagocytosis of injured cells through pathways that involve cellular membrane PS exposure (Chong et al., 2004a) and the regulation of caspases (Chong et al., 2003b,c). EPO can directly address cellular inflammation by inhibiting several pro-inflammatory cytokines, such as IL-6, tumor necrosis factor- α (TNF- α), and monocyte chemoattractant protein 1 (Chong et al., 2002c; Genc et al., 2004).

A number of trophic factors and cytokines, such as EPO, may depend upon a member of the Wnt signaling pathway. For example, new work illustrates that EPO can maintain the subcellular localization of β -catenin in the nucleus of microglia during oxidative stress that involves oxygen-glucose deprivation, suggesting that EPO protects β -catenin from degradation (Fig. 4). In addition, other components of the Wnt pathway that are utilized by EPO, such as Akt, can offer cellular

protection (Maiese et al., 2003). EPO can phosphorylate Akt and is dependent upon the activation of PI 3-K and Janus Kinase 2 (Jak2) (Witthuhn et al., 1993; Chong et al., 2002b). One of the principal pathways through which EPO prevents cellular apoptosis is through the activation of Akt (Maiese et al., 2004, 2005). During anoxia or free radical exposure, expression of the active form of Akt (phospho-Akt) is increased (Kang et al., 2003a,b). EPO can significantly enhance the activity of Akt during oxidative stress and prevent inflammatory activation of microglia (Chong et al., 2003a-c). This up-regulation of Akt activity during injury paradigms appears to be vital for EPO protection, since prevention of Akt phosphorylation blocks cellular protection by EPO (Chong et al., 2003a-c). Through the regulation of the PI 3-K/Akt dependent pathway, EPO can prevent cellular apoptosis following N-methyl-D-aspartate toxicity (Dzietko et al., 2004), hypoxia (Chong et al., 2002b), and oxidative stress (Chong et al., 2003a-c).

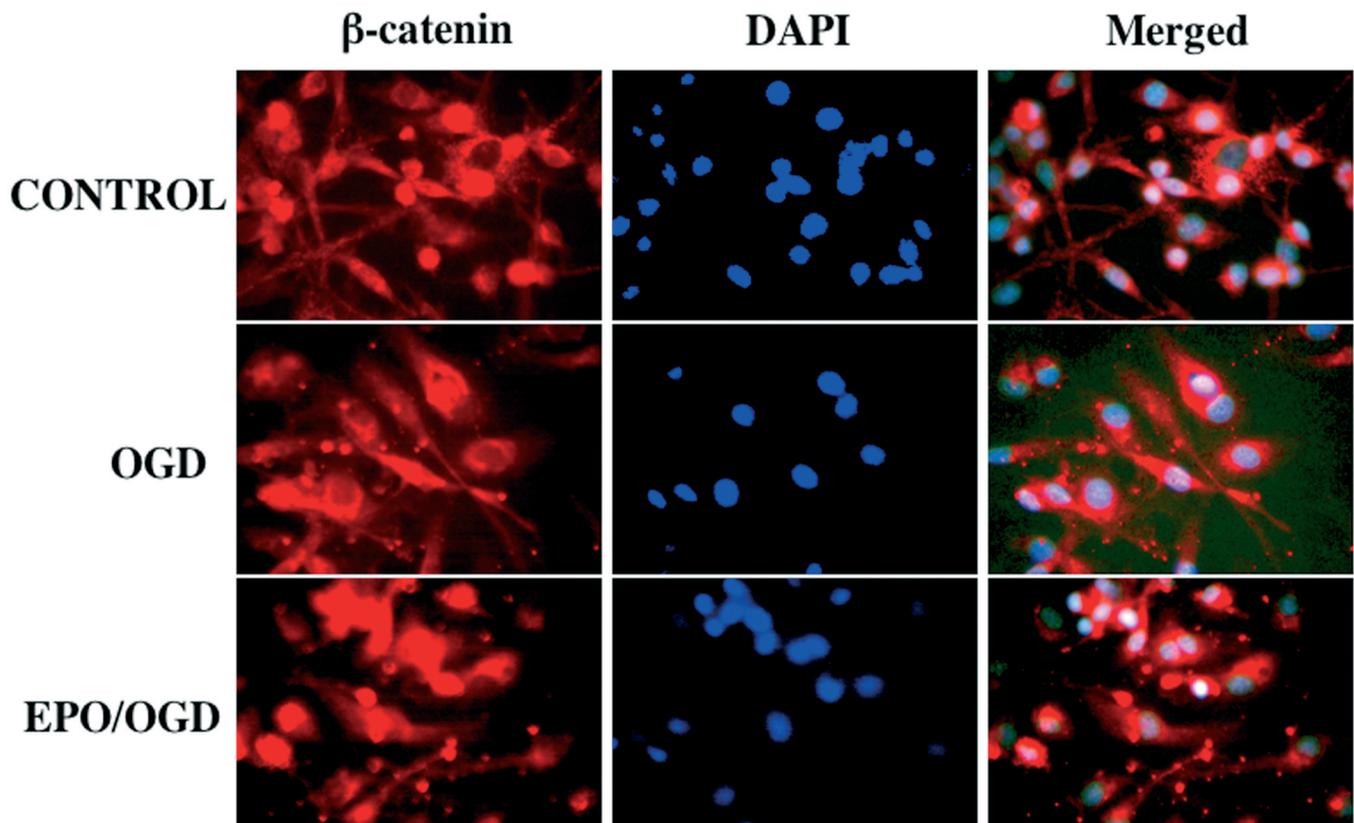


Fig. 4. Erythropoietin (EPO) employs the Wnt pathway to modulate β -catenin. The pretreatment of EPO at 1 hour before oxygen-glucose deprivation (OGD) regulates the β -catenin location in microglia following OGD. Immunohistochemical double staining for β -catenin was obtained in microglia twelve hours following an six hour period of OGD by using a primary rabbit anti- β -catenin antibody with Texas-red avidin. OGD in microglia was performed by replacing media with glucose-free HBSS containing 116 mM NaCl, 5.4 mM KCl, 0.8 mM $MgSO_4$, 1 mM NaH_2PO_4 , 0.9 mM $CaCl_2$, and 10 mg/L phenol red (pH 7.4) and cultures were maintained in an anoxic environment (95% N_2 and 5% CO_2) at 37 C for 6 hours. Cell nuclei were stained with DAPI (4'-6-diamidino-2-phenylindole). Representative images illustrate the accumulation of β -catenin in the microglial cytoplasm during OGD and the translocation of β -catenin from the cytoplasm to nuclei following the application of EPO, but the expression of β -catenin exists in both cytoplasm and nuclei without cellular translocation in control (untreated) microglia.

Conclusion

In recent years, our understanding of the Wnt-Frizzled signaling pathway has advanced at a tremendous pace. The Wnt-Frizzled signaling pathway that consists of the pathways of Wnt/ β -catenin, Wnt/ Ca^{2+} , and Wnt/planar polarity have significant roles during embryonic development, neurodegenerative disease, and cardiovascular disease. In particular, the Wnt-Frizzled signaling pathway is involved with the development of the neural plate and with the subsequent anterior-posterior extension of the neural tube. Ultimately, the Wnt-Frizzled signaling pathway leads to the development of the brain, spinal cord, and the extension of numerous sub-populations of sensory and motor neurons. If one examines the cardiovascular system, members of the Wnt family are intimately involved in vascular remodeling, cardiac development, and cardiac hypertrophy. The Wnt-Frizzled signaling pathway also controls both early and late apoptotic injury paradigms in a variety of cell populations during the development of an organism as well as during acute and chronic injury. Diseases of the nervous system, such as Alzheimer's disease or retinal degeneration, and disorders of the vascular system, such as cardiac infarction and EC injury, are closely regulated by the Wnt pathway. Furthermore, both angiogenic factors and other trophic factors rely upon members of the Wnt family. Through further identification and targeting of the critical elements that shape and control the Wnt-Frizzled signaling pathway, a greater understanding of the biological potential of Wnt-Frizzled signaling pathway can emerge for the development of new therapeutic options against neurodegenerative and vascular diseases.

Acknowledgments. This research was supported by the following grants (KM): American Heart Association (National), Bugher Foundation Award, Janssen Neuroscience Award, Johnson and Johnson Focused Investigator Award, LEARN Foundation Award, MI Life Sciences Challenge Award, and NIH NIEHS (P30 ES06639).

References

- Aberle H., Bauer A., Stappert J., Kispert A. and Kemler R. (1997). β -catenin is a target for the ubiquitin-proteasome pathway. *EMBO J.* 16, 3797-3804.
- Abu-Khalil A., Fu L., Grove E.A., Zecevic N. and Geschwind D.H. (2004). Wnt genes define distinct boundaries in the developing human brain: implications for human forebrain patterning. *J. Comp. Neurol.* 474, 276-288.
- Adler P.N., Vinson C., Park W.J., Conover S. and Klein L. (1990). Molecular structure of frizzled, a *Drosophila* tissue polarity gene. *Genetics* 126, 401-416.
- Ahmed Y., Nouri A. and Wieschaus E. (2002). *Drosophila* Apc1 and Apc2 regulate Wingless transduction throughout development. *Development* 129, 1751-1762.
- Akiyama T. (2000). Wnt/ β -catenin signaling. Cytokine growth factor Rev. 11, 273-282.
- Anderton B.H. (1999). Alzheimer's disease: clues from flies and worms. *Curr. Biol.* 9, R106-109.
- Augustine K., Liu E.T. and Sadler T.W. (1993). Antisense attenuation of Wnt-1 and Wnt-3a expression in whole embryo culture reveals roles for these genes in craniofacial, spinal cord, and cardiac morphogenesis. *Dev. Genet.* 14, 500-520.
- Axelrod J.D., Miller J.R., Shulman J.M., Moon R.T. and Perrimon N. (1998). Differential recruitment of Dishevelled provides signaling specificity in the planar cell polarity and Wingless signaling pathways. *Genes Dev.* 12, 2610-2622.
- Baker J.C., Beddington R.S. and Harland R.M. (1999). Wnt signaling in *Xenopus* embryos inhibits bmp4 expression and activates neural development. *Genes Dev.* 13, 3149-3159.
- Barandon L., Couffinhal T., Ezan J., Dufourcq P., Costet P., Alzieu P., Leroux L., Moreau C., Dare D. and Duplaa C. (2003). Reduction of infarct size and prevention of cardiac rupture in transgenic mice overexpressing FrzA. *Circulation* 108, 2282-2289.
- Bastidas F., De Calisto J. and Mayor R. (2004). Identification of neural crest competence territory: role of Wnt signaling. *Dev. Dyn.* 229, 109-117.
- Behrens J., von Kries J.P., Kuhl M., Bruhn L., Wedlich D., Grosschedl R. and Birchmeier W. (1996). Functional interaction of β -catenin with the transcription factor LEF-1. *Nature* 382, 638-642.
- Bejsovec A. (2005). Wnt pathway activation: new relations and locations. *Cell* 120, 11-14.
- Bergmann M.W., Rechner C., Freund C., Baurand A., El Jamali A. and Dietz R. (2004). Statins inhibit reoxygenation-induced cardiomyocyte apoptosis: role for glycogen synthase kinase 3 β and transcription factor β -catenin. *J. Mol. Cell. Cardiol.* 37, 681-690.
- Bhanot P., Brink M., Samos C.H., Hsieh J.C., Wang Y., Macke J.P., Andrew D., Nathans J. and Nusse R. (1996). A new member of the frizzled family from *Drosophila* functions as a Wingless receptor. *Nature*. 382, 225-230.
- Blankesteyn W.M., Gijn M.E.V., Daemen M.J.A.P., Wynshaw-Boris, Smits J.F.M. and Pratt R.E. (1999). Alterations in the cadherin-catenin complex of dishevelled-1 knockout mice lead to infarct rupture after myocardial infarction. *Circulation* 100 suppl, 156.
- Blankesteyn W.M., van Gijn M.E., Essers-Janssen Y.P., Daemen M.J. and Smits J.F. (2000). β -catenin, an inducer of uncontrolled cell proliferation and migration in malignancies, is localized in the cytoplasm of vascular endothelium during neovascularization after myocardial infarction. *Am. J. Pathol.* 157, 877-883.
- Bombeli T., Karsan A., Tait J.F. and Harlan J.M. (1997). Apoptotic vascular endothelial cells become procoagulant. *Blood* 89, 2429-2442.
- Bonvini P., An W.G., Rosolen A., Nguyen P., Trepel J., Garcia de Herreros A., Dunach M. and Neckers L.M. (2001). Geldanamycin abrogates ErbB2 association with proteasome-resistant β -catenin in melanoma cells, increases β -catenin-E-cadherin association, and decreases β -catenin-sensitive transcription. *Cancer Res.* 61, 1671-1677.
- Bose J., Gruber A.D., Helming L., Schiebe S., Wegener I., Hafner M., Beales M., Kontgen F. and Lengeling A. (2004). The phosphatidylserine receptor has essential functions during embryogenesis but not in apoptotic cell removal. *J. Biol.* 3, 15.
- Bournat J.C., Brown A.M. and Soler A.P. (2000). Wnt-1 dependent activation of the survival factor NF- κ B in PC12 cells. *J. Neurosci. Res.* 61, 21-32.

Wnt during development and demise

- Boutros M. and Mlodzik M. (1999). Dishevelled: at the crossroads of divergent intracellular signaling pathways. *Mech. Dev.* 83, 27-37.
- Boutros M., Paricio N., Strutt D.I. and Mlodzik M. (1998). Dishevelled activates JNK and discriminates between JNK pathways in planar polarity and wingless signaling. *Cell* 94, 109-118.
- Braut V., Moore R., Kutsch S., Ishibashi M., Rowitch D.H., McMahon A.P., Sommer L., Boussadia O. and Kemler R. (2001). Inactivation of the beta-catenin gene by Wnt1-Cre-mediated deletion results in dramatic brain malformation and failure of craniofacial development. *Development* 128, 1253-1264.
- Braun M.M., Etheridge A., Bernard A., Robertson C.P. and Roelink H. (2003). Wnt signaling is required at distinct stages of development for the induction of the posterior forebrain. *Development* 130, 5579-5587.
- Brigstock D.R. (2002). Regulation of angiogenesis and endothelial cell function by connective tissue growth factor (CTGF) and cysteine-rich 61 (CYR61). *Angiogenesis* 5, 153-165.
- Bronner-Fraser M. (2004). Development. Making sense of the sensory lineage. *Science* 303, 966-968.
- Burri P.H. and Djonov V. (2002). Intussusceptive angiogenesis--the alternative to capillary sprouting. *Mol. Aspects Med.* 23, S1-27.
- Cai Z., Manalo D.J., Wei G., Rodriguez E.R., Fox-Talbot K., Lu H., Zweier J.L. and Semenza G.L. (2003). Hearts from rodents exposed to intermittent hypoxia or erythropoietin are protected against ischemia-reperfusion injury. *Circulation* 108, 79-85.
- Castelo-Branco G., Wagner J., Rodriguez F.J., Kele J., Sousa K., Rawal N., Pasolli H.A., Fuchs E., Kitajewski J. and Arenas E. (2003). Differential regulation of midbrain dopaminergic neuron development by Wnt-1, Wnt-3a, and Wnt-5a. *Proc. Natl. Acad. Sci. USA* 100, 12747-12752.
- Cavallo R.A., Cox R.T., Moline M.M., Roose J., Polevoy G.A., Clevers H., Peifer M. and Bejsovec A. (1998). Drosophila Tcf and Groucho interact to repress Wingless signalling activity. *Nature* 395, 604-608.
- Chen L., Wu Q., Guo F., Xia B. and Zuo J. (2004). Expression of Dishevelled-1 in wound healing after acute myocardial infarction: possible involvement in myofibroblast proliferation and migration. *J. Cell. Mol. Med.* 8, 257-264.
- Chen S., Guttridge D.C., You Z., Zhang Z., Fribley A., Mayo M.W., Kitajewski J. and Wang C.Y. (2001). Wnt-1 signaling inhibits apoptosis by activating beta-catenin/T cell factor-mediated transcription. *J. Cell. Biol.* 152, 87-96.
- Chen T., Yang L., Irby R., Shain K.H., Wang H.G., Quackenbush J., Coppola D., Cheng J.Q. and Yeatman T.J. (2003). Regulation of caspase expression and apoptosis by adenomatous polyposis coli. *Cancer Res.* 63, 4368-4374.
- Cheng C.W., Smith S.K. and Charnock-Jones D.S. (2003). Wnt-1 signaling inhibits human umbilical vein endothelial cell proliferation and alters cell morphology. *Exp. Cell. Res.* 291, 415-425.
- Cheyette B.N. (2004). Ryk: another heretical Wnt receptor defies the canon. *Sci. STKE.* 2004, pe54.
- Chizhikov V.V. and Millen K.J. (2005). Roof plate-dependent patterning of the vertebrate dorsal central nervous system. *Dev. Biol.* 277, 287-295.
- Chong Z.Z. and Maiese K. (2004). Targeting WNT, protein kinase B, and mitochondrial membrane integrity to foster cellular survival in the nervous system. *Histol. Histopathol.* 19, 495-504.
- Chong Z.Z., Kang J.Q. and Maiese K. (2002a). Angiogenesis and plasticity: role of erythropoietin in vascular systems. *J. Hematother. Stem. Cell Res.* 11, 863-871.
- Chong Z.Z., Kang J.Q. and Maiese K. (2002b). Erythropoietin is a novel vascular protectant through activation of Akt1 and mitochondrial modulation of cysteine proteases. *Circulation* 106, 2973-2979.
- Chong Z.Z., Kang J.Q. and Maiese K. (2002c). Hematopoietic factor erythropoietin fosters neuroprotection through novel signal transduction cascades. *J. Cereb. Blood Flow Metab.* 22, 503-514.
- Chong Z.Z., Kang J.Q. and Maiese K. (2003a). Apaf-1, Bcl-xL, Cytochrome c, and Caspase-9 Form the Critical Elements for Cerebral Vascular Protection by Erythropoietin. *J. Cereb. Blood Flow Metab.* 23, 320-330.
- Chong Z.Z., Kang J.Q. and Maiese K. (2003b). Erythropoietin fosters both intrinsic and extrinsic neuronal protection through modulation of microglia, Akt1, Bad, and caspase-mediated pathways. *Br. J. Pharmacol.* 138, 1107-1118.
- Chong Z.Z., Kang J.Q. and Maiese K. (2004a). Akt1 drives endothelial cell membrane asymmetry and microglial activation through Bcl-x(L) and caspase 1, 3, and 9. *Exp. Cell Res.* 296, 196-207.
- Chong Z.Z., Kang J.Q. and Maiese K. (2004b). Essential cellular regulatory elements of oxidative stress in early and late phases of apoptosis in the central nervous system. *Antioxid. Redox. Signal.* 6, 277-287.
- Chong Z.Z., Lin S.H., Kang J.Q. and Maiese K. (2003c). Erythropoietin prevents early and late neuronal demise through modulation of Akt1 and induction of caspase 1, 3, and 8. *J. Neurosci. Res.* 71, 659-669.
- Chong Z.Z., Lin S.H., Kang J.Q. and Maiese K. (2003d). The tyrosine phosphatase SHP2 modulates MAP kinase p38 and caspase 1 and 3 to foster neuronal survival. *Cell. Mol. Neurobiol.* 23, 561-578.
- Chong Z.Z., Kang J.-Q., Li F. and Maiese K. (2005a). mGluRI targets microglial activation and selectively prevents neuronal cell engulfment through Akt and caspase dependent pathways. *Curr. Neurovasc. Res.* 2, 197-211.
- Chong Z.Z., Li F. and Maiese K. (2005b). Activating Akt and the brain's resources to drive cellular survival and prevent inflammatory injury. *Histol. Histopathol.* 20, 299-315.
- Chong Z.Z., Li F. and Maiese K. (2005c). Stress in the brain: novel cellular mechanisms of injury linked to Alzheimer's disease. *Brain. Res. Brain Res. Rev.* 49, 1-21.
- Chong Z.Z., Li F.Q. and Maiese K. (2005d). Employing new cellular therapeutic targets for Alzheimer's disease: A change for the better? *Curr. Neurovasc. Res.* 2, 55-72.
- Christian J.L., Olson D.J. and Moon R.T. (1992). Xwnt-8 modifies the character of mesoderm induced by bFGF in isolated *Xenopus* ectoderm. *EMBO. J.* 11, 33-41.
- Cohen A.W., Carbajal J.M. and Schaeffer R.C., Jr. (1999). VEGF stimulates tyrosine phosphorylation of beta-catenin and small-pore endothelial barrier dysfunction. *Am. J. Physiol.* 277, H2038-2049.
- Crowder R.J. and Freeman R.S. (2000). Glycogen synthase kinase-3 beta activity is critical for neuronal death caused by inhibiting phosphatidylinositol 3-kinase or Akt but not for death caused by nerve growth factor withdrawal. *J. Biol. Chem.* 275, 34266-34271.
- D'Amico M., Hulit J., Amanatullah D.F., Zafonte B.T., Albanese C., Bouzahzah B., Fu M., Augenlicht L.H., Donehower L.A., Takemaru K., Moon R.T., Davis R., Lisanti M.P., Shtutman M., Zhurinsky J., Ben-Ze'ev A., Troussard A.A., Dedhar S. and Pestell R.G. (2000). The integrin-linked kinase regulates the cyclin D1 gene through glycogen synthase kinase 3beta and cAMP-responsive element-binding protein-dependent pathways. *J. Biol. Chem.* 275, 32649-32657.
- Dale T.C. (1998). Signal transduction by the Wnt family of ligands.

- Biochem. J. 329, (Pt 2) 209-223.
- Danilkovitch-Miagkova A., Miagkov A., Skeel A., Nakaigawa N., Zbar B. and Leonard E.J. (2001). Oncogenic mutants of RON and MET receptor tyrosine kinases cause activation of the beta-catenin pathway. *Mol. Cell Biol.* 21, 5857-5868.
- de la Monte S.M., Chiche J., von dem Bussche A., Sanyal S., Lahousse S.A., Janssens S.P. and Bloch K.D. (2003). Nitric oxide synthase-3 overexpression causes apoptosis and impairs neuronal mitochondrial function: relevance to Alzheimer's-type neurodegeneration. *Lab. Invest.* 83, 287-298.
- Deshpande R., Inoue T., Priess J.R. and Hill R.J. (2005). *lin-17/Frizzled* and *lin-18* regulate POP-1/TCF-1 localization and cell type specification during *C. elegans* vulval development. *Dev. Biol.* 278, 118-129.
- Dombroski D., Balasubramanian K. and Schroit A.J. (2000). Phosphatidylserine expression on cell surfaces promotes antibody-dependent aggregation and thrombosis in beta2-glycoprotein I-immune mice. *J. Autoimmun.* 14, 221-229.
- Dono R., Faulhaber J., Galli A., Zuniga A., Volk T., Texido G., Zeller R. and Ehmke H. (2002). FGF2 signaling is required for the development of neuronal circuits regulating blood pressure. *Circ. Res.* 90, E5-E10.
- Doonan F. and Cotter T.G. (2004). Apoptosis: A potential therapeutic target for retinal degenerations. *Curr. Neurovasc. Res.* 1, 41-53.
- Dufourcq P., Couffinhal T., Ezan J., Barandon L., Moreau C., Daret D. and Duplaa C. (2002). FrzA, a secreted frizzled related protein, induced angiogenic response. *Circulation* 106, 3097-3103.
- Duplaa C., Jaspard B., Moreau C. and D'Amore P.A. (1999). Identification and cloning of a secreted protein related to the cysteine-rich domain of frizzled. Evidence for a role in endothelial cell growth control. *Circ. Res.* 84, 1433-1445.
- Dzietko M., Felderhoff-Mueser U., Sifringer M., Krutz B., Bittigau P., Thor F., Heumann R., Buhner C., Ikonomidou C. and Hansen H.H. (2004). Erythropoietin protects the developing brain against N-methyl-D-aspartate receptor antagonist neurotoxicity. *Neurobiol. Dis.* 15, 177-187.
- El Jamali A., Freund C., Rechner C., Scheidereit C., Dietz R. and Bergmann M.W. (2004). Reoxygenation after severe hypoxia induces cardiomyocyte hypertrophy in vitro: activation of CREB downstream of GSK3beta. *FASEB J.* 18, 1096-1098.
- Ellies D.L., Church V., Francis-West P. and Lumsden A. (2000). The WNT antagonist cSFRP2 modulates programmed cell death in the developing hindbrain. *Development* 127, 5285-5295.
- Fagotto F., Funayama N., Gluck U. and Gumbiner B.M. (1996). Binding to cadherins antagonizes the signaling activity of beta-catenin during axis formation in *Xenopus*. *J. Cell Biol.* 132, 1105-1114.
- Ferretti P. (2004). Neural stem cell plasticity: Recruitment of endogenous populations for regeneration. *Curr. Neurovasc. Res.* 1, 215-229.
- Finch P.W., He X., Kelley M.J., Uren A., Schaudies R.P., Popescu N.C., Rudikoff S., Aaronson S.A., Varmus H.E. and Rubin J.S. (1997). Purification and molecular cloning of a secreted, Frizzled-related antagonist of Wnt action. *Proc. Natl. Acad. Sci. USA.* 94, 6770-6775.
- Fukumoto S., Hsieh C.M., Maemura K., Layne M.D., Yet S.F., Lee K.H., Matsui T., Rosenzweig A., Taylor W.G., Rubin J.S., Perrella M.A. and Lee M.E. (2001). Akt participation in the Wnt signaling pathway through Dishevelled. *J. Biol. Chem.* 276, 17479-17483.
- Gabbiani G. (1998). Evolution and clinical implications of the myofibroblast concept. *Cardiovasc. Res.* 38, 545-548.
- Galceran J., Miyashita-Lin E.M., Devaney E., Rubenstein J.L. and Grosschedl R. (2000). Hippocampus development and generation of dentate gyrus granule cells is regulated by LEF1. *Development* 127, 469-482.
- Garrido J.L., Godoy J.A., Alvarez A., Bronfman M. and Inestrosa N.C. (2002). Protein kinase C inhibits amyloid beta peptide neurotoxicity by acting on members of the Wnt pathway. *Faseb. J.* 16, 1982-1984.
- Genc S., Koroglu T.F. and Genc K. (2004). Erythropoietin as a novel neuroprotectant. *Res. Neur. Neurosci.* 22, 105-119.
- Glass D.J., Bowen D.C., Stitt T.N., Radziejewski C., Bruno J., Ryan T.E., Gies D.R., Shah S., Mattsson K., Burden S.J., DiStefano P.S., Valenzuela D.M., DeChiara T.M. and Yancopoulos G.D. (1996). Agrin acts via a MuSK receptor complex. *Cell* 85, 513-523.
- Glinka A., Wu W., Delius H., Monaghan A.P., Blumenstock C. and Niehrs C. (1998). Dickkopf-1 is a member of a new family of secreted proteins and functions in head induction. *Nature* 391, 357-362.
- Golden J.A., Bracilovic A., McFadden K.A., Beesley J.S., Rubenstein J.L. and Grinspan J.B. (1999). Ectopic bone morphogenetic proteins 5 and 4 in the chicken forebrain lead to cyclopia and holoprosencephaly. *Proc. Natl. Acad. Sci. USA* 96, 2439-2444.
- Goldshmit Y., Erlich S. and Pinkas-Kramarski R. (2001). Neuregulin rescues PC12-ErbB4 cells from cell death induced by H(2)O(2). Regulation of reactive oxygen species levels by phosphatidylinositol 3-kinase. *J. Biol. Chem.* 276, 46379-46385.
- Goodwin A.M. and D'Amore P.A. (2002). Wnt signaling in the vasculature. *Angiogenesis* 5, 1-9.
- Graham N.A. and Asthagiri A.R. (2004). Epidermal growth factor-mediated T-cell factor/lymphoid enhancer factor transcriptional activity is essential but not sufficient for cell cycle progression in nontransformed mammary epithelial cells. *J. Biol. Chem.* 279, 23517-23524.
- Grilli M., Toninelli G.F., Uberti D., Spano P. and Memo M. (2003). Alzheimer's disease linking neurodegeneration with neurodevelopment. *Funct. Neurol.* 18, 145-148.
- Grimm C., Wenzel A., Groszer M., Mayser H., Seeliger M., Samardzija M., Bauer C., Gassmann M. and Reme C.E. (2002). HIF-1-induced erythropoietin in the hypoxic retina protects against light-induced retinal degeneration. *Nat. Med.* 8, 718-724.
- Grotewold L. and Ruther U. (2002a). Bmp, Fgf and Wnt signalling in programmed cell death and chondrogenesis during vertebrate limb development: the role of Dickkopf-1. *Int. J. Dev. Biol.* 46, 943-947.
- Grotewold L. and Ruther U. (2002b). The Wnt antagonist Dickkopf-1 is regulated by Bmp signaling and c-Jun and modulates programmed cell death. *EMBO. J.* 21, 966-975.
- Grove E.A. and Tole S. (1999). Patterning events and specification signals in the developing hippocampus. *Cereb. Cortex* 9, 551-561.
- Grove E.A., Tole S., Limon J., Yip L. and Ragsdale C.W. (1998). The hem of the embryonic cerebral cortex is defined by the expression of multiple Wnt genes and is compromised in Gli3-deficient mice. *Development* 125, 2315-2325.
- Habas R. and Dawid I.B. (2005). Dishevelled and Wnt signaling: is the nucleus the final frontier? *J. Biol.* 4, 2.
- Habas R., Dawid I.B. and He X. (2003). Coactivation of Rac and Rho by Wnt/Frizzled signaling is required for vertebrate gastrulation. *Genes Dev.* 17, 295-309.
- Hall A.C., Lucas F.R. and Salinas P.C. (2000). Axonal remodeling and synaptic differentiation in the cerebellum is regulated by WNT-7a signaling. *Cell* 100, 525-535.

Wnt during development and demise

- Haq S., Choukroun G., Kang Z.B., Ranu H., Matsui T., Rosenzweig A., Molkenkin J.D., Alessandrini A., Woodgett J., Hajjar R., Michael A. and Force T. (2000). Glycogen synthase kinase-3beta is a negative regulator of cardiomyocyte hypertrophy. *J. Cell Biol.* 151, 117-130.
- Haq S., Choukroun G., Lim H., Tymitz K.M., del Monte F., Gwathmey J., Grazette L., Michael A., Hajjar R., Force T. and Molkenkin J.D. (2001). Differential activation of signal transduction pathways in human hearts with hypertrophy versus advanced heart failure. *Circulation* 103, 670-677.
- Hardt S.E. and Sadoshima J. (2002). Glycogen synthase kinase-3beta: a novel regulator of cardiac hypertrophy and development. *Circ. Res.* 90, 1055-1063.
- Hari L., Braut V., Kleber M., Lee H.Y., Ille F., Leimeroth R., Paratore C., Suter U., Kemler R. and Sommer L. (2002). Lineage-specific requirements of beta-catenin in neural crest development. *J. Cell Biol.* 159, 867-880.
- Hart M., Concordet J.P., Lassot I., Albert I., del los Santos R., Durand H., Perret C., Rubinfeld B., Margottin F., Benarous R. and Polakis P. (1999). The F-box protein beta-TrCP associates with phosphorylated beta-catenin and regulates its activity in the cell. *Curr. Biol.* 9, 207-210.
- Hasegawa S., Sato T., Akazawa H., Okada H., Maeno A., Ito M., Sugitani Y., Shibata H., Miyazaki Ji J., Katsuki M., Yamauchi Y., Yamamura Ki K., Katamine S. and Noda T. (2002). Apoptosis in neural crest cells by functional loss of APC tumor suppressor gene. *Proc. Natl. Acad. Sci. USA* 99, 297-302.
- Hatsugai N., Kuroyanagi M., Yamada K., Meshi T., Tsuda S., Kondo M., Nishimura M. and Hara-Nishimura I. (2004). A plant vacuolar protease, VPE, mediates virus-induced hypersensitive cell death. *Science* 305, 855-858.
- He T.C., Sparks A.B., Rago C., Hermeking H., Zawel L., da Costa L.T., Morin P.J., Vogelstein B. and Kinzler K.W. (1998). Identification of c-MYC as a target of the APC pathway. *Science* 281, 1509-1512.
- He X.C., Zhang J., Tong W.G., Tawfik O., Ross J., Scoville D.H., Tian Q., Zeng X., He X., Wiedemann L.M., Mishina Y. and Li L. (2004). BMP signaling inhibits intestinal stem cell self-renewal through suppression of Wnt-beta-catenin signaling. *Nat. Genet.* 36, 1117-1121.
- Heisenberg C.P., Tada M., Rauch G.J., Saude L., Concha M.L., Geisler R., Stemple D.L., Smith J.C. and Wilson S.W. (2000). Silberblick/Wnt11 mediates convergent extension movements during zebrafish gastrulation. *Nature* 405, 76-81.
- Hirabayashi Y., Itoh Y., Tabata H., Nakajima K., Akiyama T., Masuyama N. and Gotoh Y. (2004). The Wnt/beta-catenin pathway directs neuronal differentiation of cortical neural precursor cells. *Development* 131, 2791-2801.
- Holthoner W., Pillinger M., Groger M., Wolff K., Ashton A.W., Albanese C., Neumeister P., Pestell R.G. and Petzelbauer P. (2002). Fibroblast growth factor-2 induces Lef/Tcf-dependent transcription in human endothelial cells. *J. Biol. Chem.* 277, 45847-45853.
- Hong J.R., Lin G.H., Lin C.J., Wang W.P., Lee C.C., Lin T.L. and Wu J.L. (2004). Phosphatidylserine receptor is required for the engulfment of dead apoptotic cells and for normal embryonic development in zebrafish. *Development* 131, 5417-5427.
- Honore S.M., Aybar M.J. and Mayor R. (2003). Sox10 is required for the early development of the prospective neural crest in *Xenopus* embryos. *Dev. Biol.* 260, 79-96.
- Hsieh J.C. (2004). Specificity of WNT-receptor interactions. *Front. Biosci.* 9, 1333-1338.
- Hsieh J.C., Rattner A., Smallwood P.M. and Nathans J. (1999). Biochemical characterization of Wnt-frizzled interactions using a soluble, biologically active vertebrate Wnt protein. *Proc. Natl. Acad. Sci. USA* 96, 3546-3551.
- Ikeda S., Kishida S., Yamamoto H., Murai H., Koyama S. and Kikuchi A. (1998). Axin, a negative regulator of the Wnt signaling pathway, forms a complex with GSK-3beta and beta-catenin and promotes GSK-3beta-dependent phosphorylation of beta-catenin. *EMBO. J.* 17, 1371-1384.
- Ikeya M., Lee S.M., Johnson J.E., McMahon A.P. and Takada S. (1997). Wnt signalling required for expansion of neural crest and CNS progenitors. *Nature* 389, 966-970.
- Ilyas M. (2005). Wnt signalling and the mechanistic basis of tumour development. *J. Pathol.* 205, 130-144.
- Inoue T., Oz H.S., Wiland D., Gharib S., Deshpande R., Hill R.J., Katz W.S. and Sternberg P.W. (2004). *C. elegans* LIN-18 is a Ryk ortholog and functions in parallel to LIN-17/Frizzled in Wnt signaling. *Cell* 118, 795-806.
- Ishikawa T., Tamai Y., Zorn A.M., Yoshida H., Seldin M.F., Nishikawa S. and Taketo M.M. (2001). Mouse Wnt receptor gene Fzd5 is essential for yolk sac and placental angiogenesis. *Development* 128, 25-33.
- Ishitani T., Ninomiya-Tsuji J. and Matsumoto K. (2003). Regulation of lymphoid enhancer factor 1/T-cell factor by mitogen-activated protein kinase-related Nemo-like kinase-dependent phosphorylation in Wnt/beta-catenin signaling. *Mol. Cell. Biol.* 23, 1379-1389.
- Itoh K., Brott B.K., Bae G.U., Ratcliffe M.J. and Sokol S.Y. (2005). Nuclear localization is required for Dishevelled function in Wnt/beta-catenin signaling. *J. Biol.* 4, 3.
- Itoh M., Takeishi Y., Nakada S., Miyamoto T., Tsunoda Y., Takahashi H., Kubota I. and Tomoike H. (2002). Long-term treatment with angiotensin II type 1 receptor antagonist, CV-11974, restores beta-catenin mRNA expression in volume-overloaded rabbit hearts. *Heart Vessels.* 17, 36-41.
- Jackson G.R., Wiedau-Pazos M., Sang T.K., Wagle N., Brown C.A., Massachi S. and Geschwind D.H. (2002). Human wild-type tau interacts with wingless pathway components and produces neurofibrillary pathology in *Drosophila*. *Neuron* 34, 509-519.
- Jamora C. and Fuchs E. (2002). Intercellular adhesion, signalling and the cytoskeleton. *Nat. Cell Biol.* 4, E101-108.
- Jennings C.G., Dyer S.M. and Burden S.J. (1993). Muscle-specific trk-related receptor with a kringle domain defines a distinct class of receptor tyrosine kinases. *Proc. Natl. Acad. Sci. USA* 90, 2895-2899.
- Jessel R., Haertel S., Socaciu C., Tykhonova S. and Diehl H.A. (2002). Kinetics of apoptotic markers in exogenously induced apoptosis of EL4 cells. *J. Cell Mol. Med.* 6, 82-92.
- Jho E.H., Zhang T., Domon C., Joo C.K., Freund J.N. and Costantini F. (2002). Wnt/beta-catenin/Tcf signaling induces the transcription of Axin2, a negative regulator of the signaling pathway. *Mol. Cell. Biol.* 22, 1172-1183.
- Jones S.E., Jomary C., Grist J., Stewart H.J. and Neal M.J. (2000). Modulated expression of secreted frizzled-related proteins in human retinal degeneration. *Neuroreport* 11, 3963-3967.
- Jordan B.K., Mohammed M., Ching S.T., Delot E., Chen X.N., Dewing P., Swain A., Rao P.N., Elejalde B.R. and Vilain E. (2001). Up-regulation of WNT-4 signaling and dosage-sensitive sex reversal in humans. *Am. J. Hum. Genet.* 68, 1102-1109.
- Kameya S., Hawes N.L., Chang B., Heckenlively J.R., Naggert J.K. and

Wnt during development and demise

- Nishina P.M. (2002). Mfrp, a gene encoding a frizzled related protein, is mutated in the mouse retinal degeneration 6. *Hum. Mol. Genet.* 11, 1879-1886.
- Kang J.Q., Chong Z.Z. and Maiese K. (2003a). Akt1 protects against inflammatory microglial activation through maintenance of membrane asymmetry and modulation of cysteine protease activity. *J. Neurosci. Res.* 74, 37-51.
- Kang J.Q., Chong Z.Z. and Maiese K. (2003b). Critical role for Akt1 in the modulation of apoptotic phosphatidylserine exposure and microglial activation. *Mol. Pharmacol.* 64, 557-569.
- Kaptanoglu E., Solaroglu I., Okutan O., Surucu H.S., Akbiyik F. and Beskonakli E. (2004). Erythropoietin exerts neuroprotection after acute spinal cord injury in rats: effect on lipid peroxidation and early ultrastructural findings. *Neurosurg. Rev.* 27, 113-120.
- Kato M. (2002). Regulation of WNT signaling molecules by retinoic acid during neuronal differentiation in NT2 cells: threshold model of WNT action. *Int. J. Mol. Med.* 10, 683-687.
- Kawakami Y., Capdevila J., Buscher D., Itoh T., Rodriguez Esteban C. and Izpisua Belmonte J.C. (2001). WNT signals control FGF-dependent limb initiation and AER induction in the chick embryo. *Cell* 104, 891-900.
- Kengaku M., Twombly V. and Tabin C. (1997). Expression of Wnt and Frizzled genes during chick limb bud development. *Cold Spring Harb. Symp. Quant. Biol.* 62, 421-429.
- Kiecker C. and Niehrs C. (2001). A morphogen gradient of Wnt/beta-catenin signalling regulates anteroposterior neural patterning in *Xenopus*. *Development* 128, 4189-4201.
- Kishida M., Hino S., Michiue T., Yamamoto H., Kishida S., Fukui A., Asashima M. and Kikuchi A. (2001). Synergistic activation of the Wnt signaling pathway by Dvl and casein kinase Iepsilon. *J. Biol. Chem.* 276, 33147-33155.
- Koyama R. and Ikegaya Y. (2004). Mossy fiber sprouting as a potential therapeutic target for epilepsy. *Curr. Neurovasc. Res.* 1, 3-10.
- Koyanagi M., Haendeler J., Badorff C., Brandes R.P., Hoffmann J., Pandur P., Zeiher A.M., Kuhl M. and Dimmeler S. (2005). Non-canonical Wnt signaling enhances differentiation of human circulating progenitor cells to cardiomyogenic cells. *J. Biol. Chem.* 280, 16838-16842.
- Kozlovsky N., Belmaker R.H. and Agam G. (2002). GSK-3 and the neurodevelopmental hypothesis of schizophrenia. *Eur. Neuropsychopharmacol.* 12, 13-25.
- Kuhl M. (2004). The WNT/calcium pathway: biochemical mediators, tools and future requirements. *Front. Biosci.* 9, 967-974.
- Kuhl M., Sheldahl L.C., Park M., Miller J.R. and Moon R.T. (2000). The Wnt/Ca²⁺ pathway: a new vertebrate Wnt signaling pathway takes shape. *Trends. Genet.* 16, 279-283.
- Kunz M., Herrmann M., Wedlich D. and Gradl D. (2004). Autoregulation of canonical Wnt signaling controls midbrain development. *Dev. Biol.* 273, 390-401.
- LaBonne C. and Bronner-Fraser M. (1998). Neural crest induction in *Xenopus*: evidence for a two-signal model. *Development* 125, 2403-2414.
- Latres E., Chiaur D.S. and Pagano M. (1999). The human F box protein beta-Trcp associates with the Cul1/Skp1 complex and regulates the stability of beta-catenin. *Oncogene* 18, 849-854.
- Lee E., Salic A. and Kirschner M.W. (2001). Physiological regulation of [beta]-catenin stability by Tcf3 and CK1epsilon. *J Cell Biol.* 154, 983-993.
- Lee H.Y., Kleber M., Hari L., Brault V., Suter U., Taketo M.M., Kemler R. and Sommer L. (2004). Instructive role of Wnt/beta-catenin in sensory fate specification in neural crest stem cells. *Science* 303, 1020-1023.
- Lee J.S., Ishimoto A. and Yanagawa S. (1999). Characterization of mouse dishevelled (Dvl) proteins in Wnt/Wingless signaling pathway. *J. Biol. Chem.* 274, 21464-21470.
- Lee S.M., Tole S., Grove E. and McMahon A.P. (2000). A local Wnt-3a signal is required for development of the mammalian hippocampus. *Development* 127, 457-467.
- Lewis J.L., Bonner J., Modrell M., Ragland J.W., Moon R.T., Dorsky R.I. and Raible D.W. (2004). Reiterated Wnt signaling during zebrafish neural crest development. *Development* 131, 1299-1308.
- Leyns L., Bouwmeester T., Kim S.H., Piccolo S. and De Robertis E.M. (1997). Frzb-1 is a secreted antagonist of Wnt signaling expressed in the Spemann organizer. *Cell* 88, 747-756.
- Li L., Yuan H., Xie W., Mao J., Caruso A.M., McMahon A., Sussman D.J. and Wu D. (1999). Dishevelled proteins lead to two signaling pathways. Regulation of LEF-1 and c-Jun N-terminal kinase in mammalian cells. *J. Biol. Chem.* 274, 129-134.
- Li F., Chong Z.Z. and Maiese K. (2004a). Erythropoietin on a tightrope: balancing neuronal and vascular protection between intrinsic and extrinsic pathways. *Neurosignals* 13, 265-289.
- Li F., Chong Z.Z. and Maiese K. (2004b). Navigating novel mechanisms of cellular plasticity with the NAD⁺ precursor and nutrient nicotinamide. *Front. Biosci.* 9, 2500-2520.
- Ligon L.A., Karki S., Tokito M. and Holzbaur E.L. (2001). Dynein binds to beta-catenin and may tether microtubules at adherens junctions. *Nat. Cell. Biol.* 3, 913-917.
- Lin S.H. and Maiese K. (2001). The metabotropic glutamate receptor system protects against ischemic free radical programmed cell death in rat brain endothelial cells. *J. Cereb. Blood Flow Metab.* 21, 262-275.
- Lisovsky M., Itoh K. and Sokol S.Y. (2002). Frizzled receptors activate a novel JNK-dependent pathway that may lead to apoptosis. *Curr. Biol.* 12, 53-58.
- Liu P., Wakamiya M., Shea M.J., Albrecht U., Behringer R.R. and Bradley A. (1999). Requirement for Wnt3 in vertebrate axis formation. *Nat. Genet.* 22, 361-365.
- Longo K.A., Kennell J.A., Ochocinska M.J., Ross S.E., Wright W.S. and MacDougald O.A. (2002). Wnt signaling protects 3T3-L1 preadipocytes from apoptosis through induction of insulin-like growth factors. *J. Biol. Chem.* 277, 38239-38244.
- Lu W., Yamamoto V., Ortega B. and Baltimore D. (2004). Mammalian Ryk is a Wnt coreceptor required for stimulation of neurite outgrowth. *Cell* 119, 97-108.
- Lustig B. and Behrens J. (2003). The Wnt signaling pathway and its role in tumor development. *J. Cancer Res. Clin. Oncol.* 129, 199-221.
- Lustig B., Jerchow B., Sachs M., Weiler S., Pietsch T., Karsten U., van de Wetering M., Clevers H., Schlag P.M., Birchmeier W. and Behrens J. (2002). Negative feedback loop of Wnt signaling through upregulation of conductin/axin2 in colorectal and liver tumors. *Mol. Cell Biol.* 22, 1184-1193.
- Machon O., van den Bout C.J., Backman M., Kemler R. and Krauss S. (2003). Role of beta-catenin in the developing cortical and hippocampal neuroepithelium. *Neuroscience.* 122, 129-143.
- Madaio M.P., Fabbi M., Tiso M., Daga A. and Puccetti A. (1996). Spontaneously produced anti-DNA/DNase I autoantibodies modulate nuclear apoptosis in living cells. *Eur. J. Immunol.* 26, 3035-3041.

Wnt during development and demise

- Maiese K. (2001). The dynamics of cellular injury: transformation into neuronal and vascular protection. *Histol. Histopathol.* 16, 633-644.
- Maiese K. and Chong Z.Z. (2003). Nicotinamide: necessary nutrient emerges as a novel cytoprotectant for the brain. *Trends. Pharmacol. Sci.* 24, 228-232.
- Maiese K. and Chong Z.Z. (2004). Insights into oxidative stress and potential novel therapeutic targets for Alzheimer disease. *Restor. Neurol. Neurosci.* 22, 87-104.
- Maiese K., Chong Z.Z. and Kang J. (2003). Transformation into treatment: Novel therapeutics that begin within the cell. In: *Neuronal and vascular plasticity: Elucidating basic cellular mechanisms for future therapeutic discovery.* Maiese K. (ed). Kluwer Academic Publishers: Norwell, MA. pp 1-26.
- Maiese K., Li F. and Chong Z.Z. (2004). Erythropoietin in the brain: can the promise to protect be fulfilled? *Trends. Pharmacol. Sci.* 25, 577-583.
- Maiese K., Li F. and Chong Z.Z. (2005). New avenues of exploration for erythropoietin. *JAMA* 293, 90-95.
- Maiese K. and Vincent A.M. (2000). Membrane asymmetry and DNA degradation: functionally distinct determinants of neuronal programmed cell death. *J. Neurosci. Res.* 59, 568-580.
- Mao C., Malek O.T., Pueyo M.E., Steg P.G. and Soubrier F. (2000). Differential expression of rat frizzled-related frzb-1 and frizzled receptor fz1 and fz2 genes in the rat aorta after balloon injury. *Arterioscler. Thromb. Vasc. Biol.* 20, 43-51.
- Mao J., Wang J., Liu B., Pan W., Farr G.H., 3rd Flynn C., Yuan H., Takada S., Kimelman D., Li L. and Wu D. (2001). Low-density lipoprotein receptor-related protein-5 binds to Axin and regulates the canonical Wnt signaling pathway. *Mol. Cell.* 7, 801-809.
- Marvin M.J., Di Rocco G., Gardiner A., Bush S.M. and Lassar A.B. (2001). Inhibition of Wnt activity induces heart formation from posterior mesoderm. *Genes Dev.* 15, 316-327.
- Mattson M.P. (2004). Pathways towards and away from Alzheimer's disease. *Nature* 430, 631-639.
- Mayr T., Deutsch U., Kuhl M., Drexler H.C., Lottspeich F., Deutzmann R., Wedlich D. and Risau W. (1997). Fritz: a secreted frizzled-related protein that inhibits Wnt activity. *Mech. Dev.* 63, 109-125.
- Melkonyan H.S., Chang W.C., Shapiro J.P., Mahadevappa M., Fitzpatrick P.A., Kiefer M.C., Tomei L.D. and Umansky S.R. (1997). SARRPs: a family of secreted apoptosis-related proteins. *Proc. Natl. Acad. Sci. USA* 94, 13636-13641.
- Melton K.R., Lulianella A. and Trainor P.A. (2004). Gene expression and regulation of hindbrain and spinal cord development. *Front. Biosci.* 9, 117-138.
- Michiue T., Fukui A., Yukita A., Sakurai K., Danno H., Kikuchi A. and Asashima M. (2004). Xlax, an inhibitor of the canonical Wnt pathway, is required for anterior neural structure formation in *Xenopus*. *Dev. Dyn.* 230, 79-90.
- Miller J.R. (2002). The Wnts. *Genome Biol.* 3, REVIEWS3001.
- Mirkovic I., Charish K., Gorski S.M., McKnight K. and Verheyen E.M. (2002). *Drosophila nemo* is an essential gene involved in the regulation of programmed cell death. *Mech. Dev.* 119, 9-20.
- Miyaoka T., Seno H. and Ishino H. (1999). Increased expression of Wnt-1 in schizophrenic brains. *Schizophr. Res.* 38, 1-6.
- Montague J.W., Hughes F., Jr. and Cidlowski J.A. (1997). Native recombinant cyclophilins A, B, and C degrade DNA independently of peptidylprolyl cis-trans-isomerase activity. Potential roles of cyclophilins in apoptosis. *J. Biol. Chem.* 272, 6677-6684.
- Moon C., Krawczyk M., Ahn D., Ahmet I., Paik D., Lakatta E.G. and Talan M.I. (2003). Erythropoietin reduces myocardial infarction and left ventricular functional decline after coronary artery ligation in rats. *Proc. Natl. Acad. Sci. USA* 100, 11612-11617.
- Moon R.T., Campbell R.M., Christian J.L., McGrew L.L., Shih J. and Fraser S. (1993). Xwnt-5A: a maternal Wnt that affects morphogenetic movements after overexpression in embryos of *Xenopus laevis*. *Development* 119, 97-111.
- Moriguchi T., Kawachi K., Kamakura S., Masuyama N., Yamanaka H., Matsumoto K., Kikuchi A. and Nishida E. (1999). Distinct domains of mouse dishevelled are responsible for the c-Jun N-terminal kinase/stress-activated protein kinase activation and the axis formation in vertebrates. *J. Biol. Chem.* 274, 30957-30962.
- Moulin N. and Widmann C. (2004). Islet-Brain (IB)/JNK-interacting proteins (JIPs): Future targets for the treatment of neurodegenerative diseases? *Curr. Neurovasc. Res.* 1, 111-127.
- Mudher A., Chapman S., Richardson J., Asuni A., Gibb G., Pollard C., Killick R., Iqbal T., Raymond L., Varndell I., Sheppard P., Makoff A., Gower E., Soden P.E., Lewis P., Murphy M., Golde T.E., Rupniak H.T., Anderton B.H. and Lovestone S. (2001). Dishevelled regulates the metabolism of amyloid precursor protein via protein kinase C/mitogen-activated protein kinase and c-Jun terminal kinase. *J. Neurosci.* 21, 4987-4995.
- Munemitsu S., Albert I., Souza B., Rubinfeld B. and Polakis P. (1995). Regulation of intracellular beta-catenin levels by the adenomatous polyposis coli (APC) tumor-suppressor protein. *Proc. Natl. Acad. Sci. USA* 92, 3046-3050.
- Muroyama Y., Fujihara M., Ikeya M., Kondoh H. and Takada S. (2002). Wnt signaling plays an essential role in neuronal specification of the dorsal spinal cord. *Genes. Dev.* 16, 548-553.
- Muroyama Y., Kondoh H. and Takada S. (2004). Wnt proteins promote neuronal differentiation in neural stem cell culture. *Biochem. Biophys. Res. Commun.* 313, 915-921.
- Nacher V., Carretero A., Navarro M., Armengol C., Lombart C., Blasi J. and Ruberte J. (2005). beta-Catenin expression during vascular development and degeneration of avian mesonephros. *J. Anat.* 206, 165-174.
- Nelson W.J. and Nusse R. (2004). Convergence of Wnt, beta-catenin, and cadherin pathways. *Science* 303, 1483-1487.
- Niemann S., Zhao C., Pascu F., Stahl U., Aulepp U., Niswander L., Weber J.L. and Muller U. (2004). Homozygous WNT3 mutation causes tetra-amelia in a large consanguineous family. *Am. J. Hum. Genet.* 74, 558-563.
- Noria S., Cowan D.B., Gottlieb A.I. and Langille B.L. (1999). Transient and steady-state effects of shear stress on endothelial cell adherens junctions. *Circ. Res.* 85, 504-514.
- Nusse R. (1999). WNT targets. Repression and activation. *Trends Genet.* 15, 1-3.
- Nusse R. and Varmus H.E. (1982). Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome. *Cell* 31, 99-109.
- Nusse R. and Varmus H.E. (1992). Wnt genes. *Cell* 69, 1073-1087.
- Pandey S., Walker P.R. and Sikorska M. (1997). Identification of a novel 97 kDa endonuclease capable of internucleosomal DNA cleavage. *Biochemistry* 36, 711-720.
- Panhuysen M., Vogt Weisenhorn D.M., Blanquet V., Brodski C., Heinzmann U., Beisker W. and Wurst W. (2004). Effects of Wnt1 signaling on proliferation in the developing mid-/hindbrain region. *Mol. Cell. Neurosci.* 26, 101-111.
- Papkoff J. and Aikawa M. (1998). WNT-1 and HGF regulate GSK3 beta

- activity and beta-catenin signaling in mammary epithelial cells. *Biochem. Biophys. Res. Commun.* 247, 851-858.
- Park M., Wu X., Golden K., Axelrod J.D. and Bodmer R. (1996). The wingless signaling pathway is directly involved in *Drosophila* heart development. *Dev. Biol.* 177, 104-116.
- Parsa C.J., Matsumoto A., Kim J., Riel R.U., Pascal L.S., Walton G.B., Thompson R.B., Petrofski J.A., Annex B.H., Stamler J.S. and Koch W.J. (2003). A novel protective effect of erythropoietin in the infarcted heart. *J. Clin. Invest.* 112, 999-1007.
- Parsa C.J., Kim J., Riel R.U., Pascal L.S., Thompson R.B., Petrofski J.A., Matsumoto A., Stamler J.S. and Koch W.J. (2004). Cardioprotective effects of erythropoietin in the reperfused ischemic heart: a potential role for cardiac fibroblasts. *J. Biol. Chem.* 279, 20655-20662.
- Patapoutian A. and Reichardt L.F. (2000). Roles of Wnt proteins in neural development and maintenance. *Curr. Opin. Neurobiol.* 10, 392-399.
- Pathy L. (2000). The WIF module. *Trends Biochem. Sci.* 25, 12-13.
- Pinson K.I., Brennan J., Monkley S., Avery B.J. and Skarnes W.C. (2000). An LDL-receptor-related protein mediates Wnt signalling in mice. *Nature* 407, 535-538.
- Playford M.P., Bicknell D., Bodmer W.F. and Macaulay V.M. (2000). Insulin-like growth factor 1 regulates the location, stability, and transcriptional activity of beta-catenin. *Proc. Natl. Acad. Sci. USA.* 97, 12103-12108.
- Price R.M., Tulsyan N., Dermody J.J., Schwalb M., Soteropoulos P. and Castronuovo J.J. Jr. (2004). Gene expression after crush injury of human saphenous vein: using microarrays to define the transcriptional profile. *J. Am. Coll. Surg.* 199, 411-418.
- Pugazhenthii S., Nesterova A., Jambal P., Audesirk G., Kern M., Cabell L., Eves E., Rosner M.R., Boxer L.M. and Reusch J.E. (2003). Oxidative stress-mediated down-regulation of bcl-2 promoter in hippocampal neurons. *J. Neurochem.* 84, 982-996.
- Rattner A., Hsieh J.C., Smallwood P.M., Gilbert D.J., Copeland N.G., Jenkins N.A. and Nathans J. (1997). A family of secreted proteins contains homology to the cysteine-rich ligand-binding domain of frizzled receptors. *Proc. Natl. Acad. Sci. USA* 94, 2859-2863.
- Rehn M. and Pihlajaniemi T. (1995). Identification of three N-terminal ends of type XVIII collagen chains and tissue-specific differences in the expression of the corresponding transcripts. The longest form contains a novel motif homologous to rat and *Drosophila* frizzled proteins. *J. Biol. Chem.* 270, 4705-4711.
- Reya T., Duncan A.W., Ailles L., Domen J., Scherer D.C., Willert K., Hintz L., Nusse R. and Weissman I.L. (2003). A role for Wnt signalling in self-renewal of haematopoietic stem cells. *Nature* 423, 409-414.
- Rhee C.S., Sen M., Lu D., Wu C., Leoni L., Rubin J., Corr M. and Carson D.A. (2002). Wnt and frizzled receptors as potential targets for immunotherapy in head and neck squamous cell carcinomas. *Oncogene* 21, 6598-6605.
- Ribatti D., Presta M., Vacca A., Ria R., Giuliani R., Dell'Era P., Nico B., Roncali L. and Dammacco F. (1999). Human erythropoietin induces a pro-angiogenic phenotype in cultured endothelial cells and stimulates neovascularization in vivo. *Blood* 93, 2627-2636.
- Risau W. (1997). Mechanisms of angiogenesis. *Nature* 386, 671-674.
- Robitaille J., MacDonald M.L., Kaykas A., Sheldahl L.C., Zeisler J., Dube M.P., Zhang L.H., Singaraja R.R., Guernsey D.L., Zheng B., Siebert L.F., Hoskin-Mott A., Trese M.T., Pimstone S.N., Shastri B.S., Moon R.T., Hayden M.R., Goldberg Y.P. and Samuels M.E. (2002). Mutant frizzled-4 disrupts retinal angiogenesis in familial exudative vitreoretinopathy. *Nat. Genet.* 32, 326-330.
- Rodova M., Islam M.R., Maser R.L. and Calvet J.P. (2002). The polycystic kidney disease-1 promoter is a target of the beta-catenin/T-cell factor pathway. *J. Biol. Chem.* 277, 29577-29583.
- Roose J., Molenaar M., Peterson J., Hurenkamp J., Brantjes H., Moerer P., van de Wetering M., Destree O. and Clevers H. (1998). The *Xenopus* Wnt effector XTcf-3 interacts with Groucho-related transcriptional repressors. *Nature* 395, 608-612.
- Ross S.E., Hemati N., Longo K.A., Bennett C.N., Lucas P.C., Erickson R.L. and MacDougald O.A. (2000). Inhibition of adipogenesis by Wnt signaling. *Science* 289, 950-953.
- Rosso S.B., Sussman D., Wynshaw-Boris A. and Salinas P.C. (2005). Wnt signaling through Dishevelled, Rac and JNK regulates dendritic development. *Nat. Neurosci.* 8, 34-42.
- Roura S., Miravet S., Piedra J., Garcia de Herreros A. and Dunach M. (1999). Regulation of E-cadherin/Catenin association by tyrosine phosphorylation. *J. Biol. Chem.* 274, 36734-36740.
- Sadot E., Simcha I., Shtutman M., Ben-Ze'ev A. and Geiger B. (1998). Inhibition of beta-catenin-mediated transactivation by cadherin derivatives. *Proc. Natl. Acad. Sci. USA* 95, 15339-15344.
- Sakamaki K. (2004). Regulation of endothelial cell death and its role in angiogenesis and vascular regression. *Curr. Neurovasc. Res.* 1, 305-315.
- Salinas M., Diaz R., Abraham N.G., Ruiz de Galarreta C.M. and Cuadrado A. (2003). Nerve growth factor protects against 6-hydroxydopamine-induced oxidative stress by increasing expression of heme oxygenase-1 in a phosphatidylinositol 3-kinase-dependent manner. *J. Biol. Chem.* 278, 13898-13904.
- Salinas P.C. (1999). Wnt factors in axonal remodelling and synaptogenesis. *Biochem. Soc. Symp.* 65, 101-109.
- Savage M.J., Trusko S.P., Howland D.S., Pinsker L.R., Mistretta S., Reaume A.G., Greenberg B.D., Siman R. and Scott R.W. (1998). Turnover of amyloid beta-protein in mouse brain and acute reduction of its level by phorbol ester. *J. Neurosci.* 18, 1743-1752.
- Schneider S., Buchert M., Georgiev O., Catimel B., Halford M., Stacker S.A., Baechi T., Moelling K. and Hovens C.M. (1999). Mutagenesis and selection of PDZ domains that bind new protein targets. *Nat. Biotechnol.* 17, 170-175.
- Schneider V.A. and Mercola M. (2001). Wnt antagonism initiates cardiogenesis in *Xenopus laevis*. *Genes Dev.* 15, 304-315.
- Schumann H., Holtz J., Zerkowski H.R. and Hatzfeld M. (2000). Expression of secreted frizzled related proteins 3 and 4 in human ventricular myocardium correlates with apoptosis related gene expression. *Cardiovasc. Res.* 45, 720-728.
- Sharma S.K. and Ebadi M. (2003). Metallothionein attenuates 3-morpholinosydnonimine (SIN-1)-induced oxidative stress in dopaminergic neurons. *Antioxid. Redox. Signal.* 5, 251-264.
- Shimogori T., VanSant J., Paik E. and Grove E.A. (2004). Members of the Wnt, Fz, and Frp gene families expressed in postnatal mouse cerebral cortex. *J. Comp. Neurol.* 473, 496-510.
- Shou J., Ali-Osman F., Multani A.S., Pathak S., Fedi P. and Srivenugopal K.S. (2002). Human Dkk-1, a gene encoding a Wnt antagonist, responds to DNA damage and its overexpression sensitizes brain tumor cells to apoptosis following alkylation damage of DNA. *Oncogene* 21, 878-889.
- Shtutman M., Zhurinsky J., Simcha I., Albanese C., D'Amico M., Pestell R. and Ben-Ze'ev A. (1999). The cyclin D1 gene is a target of the beta-catenin/LEF-1 pathway. *Proc. Natl. Acad. Sci. USA* 96, 5522-

Wnt during development and demise

- 5527.
- Simcha I., Shtutman M., Salomon D., Zhurinsky J., Sadot E., Geiger B. and Ben-Ze'ev A. (1998). Differential nuclear translocation and transactivation potential of beta-catenin and plakoglobin. *J. Cell Biol.* 141, 1433-1448.
- Slusarski D.C., Corces V.G. and Moon R.T. (1997). Interaction of Wnt and a Frizzled homologue triggers G-protein-linked phosphatidylinositol signalling. *Nature* 390, 410-413.
- Smalley M.J. and Dale T.C. (1999). Wnt signalling in mammalian development and cancer. *Cancer Metastasis Rev.* 18, 215-230.
- Smith W.C. and Harland R.M. (1991). Injected Xwnt-8 RNA acts early in *Xenopus* embryos to promote formation of a vegetal dorsalizing center. *Cell* 67, 753-765.
- Sokol S., Christian J.L., Moon R.T. and Melton D.A. (1991). Injected Wnt RNA induces a complete body axis in *Xenopus* embryos. *Cell* 67, 741-752.
- Song L. and Fricker L.D. (1997). Cloning and expression of human carboxypeptidase Z, a novel metallo-carboxypeptidase. *J. Biol. Chem.* 272, 10543-10550.
- Soriano S., Kang D.E., Fu M., Pestell R., Chevallier N., Zheng H. and Koo E.H. (2001). Presenilin 1 negatively regulates beta-catenin/T cell factor/lymphoid enhancer factor-1 signaling independently of beta-amyloid precursor protein and notch processing. *J. Cell Biol.* 152, 785-794.
- Stark K., Vainio S., Vassileva G. and McMahon A.P. (1994). Epithelial transformation of metanephric mesenchyme in the developing kidney regulated by Wnt-4. *Nature* 372, 679-683.
- Su F., Overholtzer M., Besser D. and Levine A.J. (2002). WISP-1 attenuates p53-mediated apoptosis in response to DNA damage through activation of the Akt kinase. *Genes Dev.* 16, 46-57.
- Sun X.M. and Cohen G.M. (1994). Mg⁽²⁺⁾-dependent cleavage of DNA into kilobase pair fragments is responsible for the initial degradation of DNA in apoptosis. *J. Biol. Chem.* 269, 14857-14860.
- Tada M. and Smith J.C. (2000). Xwnt11 is a target of *Xenopus* Brachyury: regulation of gastrulation movements via Dishevelled, but not through the canonical Wnt pathway. *Development* 127, 2227-2238.
- Tamai K., Semenov M., Kato Y., Spokony R., Liu C., Katsuyama Y., Hess F., Saint-Jeannet J.P. and He X. (2000). LDL-receptor-related proteins in Wnt signal transduction. *Nature* 407, 530-535.
- Tanaka H., Shan W., Phillips G.R., Arndt K., Bozdagi O., Shapiro L., Huntley G.W., Benson D.L. and Colman D.R. (2000). Molecular modification of N-cadherin in response to synaptic activity. *Neuron* 25, 93-107.
- Tetsu O. and McCormick F. (1999). Beta-catenin regulates expression of cyclin D1 in colon carcinoma cells. *Nature* 398, 422-426.
- Tolwinski N.S., Wehrli M., Rives A., Erdeniz N., DiNardo S. and Wieschaus E. (2003). Wg/Wnt signal can be transmitted through arrow/LRP5,6 and Axin independently of Zw3/Gsk3beta activity. *Dev. Cell.* 4, 407-418.
- Torriglia A., Chaudun E., Chany-Fournier F., Jeanny J.C., Courtois Y. and Counis M.F. (1995). Involvement of DNase II in nuclear degeneration during lens cell differentiation. *J. Biol. Chem.* 270, 28579-28585.
- Torriglia A., Chaudun E., Courtois Y. and Counis M.F. (1997). On the use of Zn²⁺ to discriminate endonucleases activated during apoptosis. *Biochimie* 79, 435-438.
- van Gijn M.E., Blankesteyn W.M., Smits J.F., Hierck B. and Gittenberger-de Groot A.C. (2001). Frizzled 2 is transiently expressed in neural crest-containing areas during development of the heart and great arteries in the mouse. *Anat. Embryol. (Berl.)* 203, 185-192.
- van Gijn M.E., Daemen M.J., Smits J.F. and Blankesteyn W.M. (2002). The wnt-frizzled cascade in cardiovascular disease. *Cardiovasc. Res.* 55, 16-24.
- Veeman M.T., Axelrod J.D. and Moon R.T. (2003). A second canon. Functions and mechanisms of beta-catenin-independent Wnt signaling. *Dev. Cell.* 5, 367-377.
- Vincent A.M. and Maiese K. (1999a). Direct temporal analysis of apoptosis induction in living adherent neurons. *J. Histochem. Cytochem.* 47, 661-672.
- Vincent A.M. and Maiese K. (1999b). Nitric oxide induction of neuronal endonuclease activity in programmed cell death. *Exp. Cell Res.* 246, 290-300.
- Vinson C.R., Conover S. and Adler P.N. (1989). A *Drosophila* tissue polarity locus encodes a protein containing seven potential transmembrane domains. *Nature* 338, 263-264.
- Wagner U., Brownlees J., Irving N.G., Lucas F.R., Salinas P.C. and Miller C.C. (1997). Overexpression of the mouse dishevelled-1 protein inhibits GSK-3beta-mediated phosphorylation of tau in transfected mammalian cells. *FEBS Lett.* 411, 369-372.
- Wang H.Y. and Malbon C.C. (2003). Wnt signaling, Ca²⁺, and cyclic GMP: visualizing Frizzled functions. *Science* 300, 1529-1530.
- Wang H.Y. and Malbon C.C. (2004). Wnt-frizzled signaling to G-protein-coupled effectors. *Cell Mol Life Sci.* 61, 69-75.
- Wang J.Y., Shum A.Y. and Ho Y.J. (2003). Oxidative neurotoxicity in rat cerebral cortex neurons: synergistic effects of H₂O₂ and NO on apoptosis involving activation of p38 mitogen-activated protein kinase and caspase-3. *J. Neurosci. Res.* 72, 508-519.
- Wang S., Krinks M., Lin K., Luyten F.P. and Moos M., Jr. (1997a). Frzb, a secreted protein expressed in the Spemann organizer, binds and inhibits Wnt-8. *Cell* 88, 757-766.
- Wang S., Krinks M. and Moos M. Jr. (1997b). Frzb-1, an antagonist of Wnt-1 and Wnt-8, does not block signaling by Wnts -3A, -5A, or -11. *Biochem. Biophys. Res. Commun.* 236, 502-504.
- Wang X., Xiao Y., Mou Y., Zhao Y., Blankesteyn W.M. and Hall J.L. (2002). A role for the beta-catenin/T-cell factor signaling cascade in vascular remodeling. *Circ. Res.* 90, 340-347.
- Wang Y., Macke J.P., Abella B.S., Andreasson K., Worley P., Gilbert D.J., Copeland N.G., Jenkins N.A. and Nathans J. (1996). A large family of putative transmembrane receptors homologous to the product of the *Drosophila* tissue polarity gene frizzled. *J. Biol. Chem.* 271, 4468-4476.
- Weber J.T. (2004). Calcium homeostasis following traumatic neuronal injury. *Curr. Neurovasc. Res.* 1, 151-171.
- Wehrli M., Dougan S.T., Caldwell K., O'Keefe L., Schwartz S., Vaizel-Ohayon D., Schejter E., Tomlinson A. and DiNardo S. (2000). Arrow encodes an LDL-receptor-related protein essential for Wingless signalling. *Nature* 407, 527-530.
- Weitzman J.B. (2005). Dishevelled nuclear shuttling. *J. Biol.* 4, 1.
- Wharton K.A. Jr. (2003). Runnin' with the Dvl: proteins that associate with Dsh/Dvl and their significance to Wnt signal transduction. *Dev. Biol.* 253, 1-17.
- Wheeler G.N. and Hoppler S. (1999). Two novel *Xenopus* frizzled genes expressed in developing heart and brain. *Mech. Dev.* 86, 203-207.
- Willert K., Brown J.D., Danenberg E., Duncan A.W., Weissman I.L.,

Wnt during development and demise

- Reya T., Yates J.R., 3rd and Nusse R. (2003). Wnt proteins are lipid-modified and can act as stem cell growth factors. *Nature* 423, 448-452.
- Wilson C., Goberdhan D.C. and Steller H. (1993). Dror, a potential neurotrophic receptor gene, encodes a Drosophila homolog of the vertebrate Ror family of Trk-related receptor tyrosine kinases. *Proc. Natl. Acad. Sci. USA* 90, 7109-7113.
- Winklbauer R., Medina A., Swain R.K. and Steinbeisser H. (2001). Frizzled-7 signalling controls tissue separation during Xenopus gastrulation. *Nature* 413, 856-860.
- Winston J.T., Strack P., Beer-Romero P., Chu C.Y., Elledge S.J. and Harper J.W. (1999). The SCFbeta-TRCP-ubiquitin ligase complex associates specifically with phosphorylated destruction motifs in IkkappaBalph and beta-catenin and stimulates IkkappaBalph ubiquitination in vitro. *Genes Dev.* 13, 270-283.
- Witthuhn B.A., Quelle F.W., Silvennoinen O., Yi T., Tang B., Miura O. and Ihle J.N. (1993). JAK2 associates with the erythropoietin receptor and is tyrosine phosphorylated and activated following stimulation with erythropoietin. *Cell* 74, 227-236.
- Witting A., Muller P., Herrmann A., Kettenmann H. and Nolte C. (2000). Phagocytic clearance of apoptotic neurons by Microglia/Brain macrophages in vitro: involvement of lectin-, integrin-, and phosphatidylserine-mediated recognition. *J. Neurochem.* 75, 1060-1070.
- Wodarz A. and Nusse R. (1998). Mechanisms of Wnt signaling in development. *Annu. Rev. Cell Dev. Biol.* 14, 59-88.
- Wolda S.L., Moody C.J. and Moon R.T. (1993). Overlapping expression of Xwnt-3A and Xwnt-1 in neural tissue of Xenopus laevis embryos. *Dev. Biol.* 155, 46-57.
- Wong G.T., Gavin B.J. and McMahon A.P. (1994). Differential transformation of mammary epithelial cells by Wnt genes. *Mol. Cell Biol.* 14, 6278-6286.
- Wright M., Aikawa M., Szeto W. and Papkoff J. (1999). Identification of a Wnt-responsive signal transduction pathway in primary endothelial cells. *Biochem. Biophys. Res. Commun.* 263, 384-388.
- Xu Q., D'Amore P.A. and Sokol S.Y. (1998). Functional and biochemical interactions of Wnts with FrZA, a secreted Wnt antagonist. *Development* 125, 4767-4776.
- Yamamoto H., Kishida S., Kishida M., Ikeda S., Takada S. and Kikuchi A. (1999). Phosphorylation of axin, a Wnt signal negative regulator, by glycogen synthase kinase-3beta regulates its stability. *J. Biol. Chem.* 274, 10681-10684.
- Yan W., Sheng N., Seto M., Morser J. and Wu Q. (1999). Corin, a mosaic transmembrane serine protease encoded by a novel cDNA from human heart. *J. Biol. Chem.* 274, 14926-14935.
- Yeo W. and Gautier J. (2004). Early neural cell death: dying to become neurons. *Dev. Biol.* 274, 233-244.
- Yoshikawa S., McKinnon R.D., Kokel M. and Thomas J.B. (2003). Wnt-mediated axon guidance via the Drosophila Derailed receptor. *Nature* 422, 583-588.
- You Z., Saims D., Chen S., Zhang Z., Guttridge D.C., Guan K.L., MacDougald O.A., Brown A.M., Evan G., Kitajewski J. and Wang C.Y. (2002). Wnt signaling promotes oncogenic transformation by inhibiting c-Myc-induced apoptosis. *J. Cell Biol.* 157, 429-440.

Accepted July 22, 2005