Summary. There has been a long persisting dilemma about potential ovarian stem cells in adult mammalian ovaries, including human, and now there is steadily increasing experimental evidence on their existence. After some previous indirect evidence about the presence of stem cells in adult mouse ovaries, an important breakthrough was made by Zou and his co-workers who successfully established long-persisting pluripotent/multipotent ovarian stem cell lines in neonatal and adult mice, and were followed by some other important studies in mouse and human. Moreover, oocyte-like cells can be developed in vitro from pluripotent stem cells of different origins (embryonic stem cells, induced pluripotent stem cells, fetal skin stem cells, pancreatic stem cells). The aim of this article is to elucidate the fast growing new knowledge on the ovarian stem cells and potential in vitro oogenesis in mammals.

Key words: Germinal stem cell, Oocyte, Oogenesis in vitro, Ovary

Introduction

Currently there is steadily increasing experimental evidence on the existence of pluripotent/multipotent stem cells in neonatal and adult mammalian ovaries. There is also increasing evidence about the potential in vitro development of oocyte-like cells from pluripotent stem cells of different origins, e.g., stem cells isolated from the porcine fetal skin and induced pluripotent stem cells in human. The aim of this article is to review the fast growing new knowledge on the ovarian stem cells and potential in vitro oogenesis in mammals.

Stem cells in mammalian fetal ovaries

Primordial germ cells (PGCs) are undifferentiated cells in developing fetuses which give rise to gametes, oocytes and spermatozoa, and contribute to new life in the next generation. They arise outside the genital ridge region, and are first identifiable in the human embryo at about 3 weeks in the yolk sac epithelium near the base of the developing alantois. Then the PGC population, expanded by mitosis, migrates by amoeboid movement to the connective tissue of the hind gut and from there into the gut mesentery. From about 30 days after fertilization the majority of cells pass into the region of the developing kidneys, and then into the adjacent gonadal primordia where they join the cells of the sex and medullary cords (Johnson and Everitt, 2000). They arise in the fetal ovaries as small clusters of 40-50 alkaline phosphatase-positive cells (Sabour et al., 2011). Their development, differentiation, and survival (apoptosis) in the fetal gonads are strictly controlled regarding their genetic and epigenetic factors, and are regulated by a combination of different growth factors and other molecular regulators known as the germ cell niche. By establishing long term cell cultures of human fetal ovaries it has been confirmed that bone morphogenetic protein (BMP) is developmentally regulated and one of the key regulators of the PGCs’ fate by promoting their apoptosis (Childs et al., 2010). Human PGCs can develop into pluripotent stem cells such as embryonal carcinoma cells (ECCs) and embryonic germ cells (EGCs). PGCs express most, but not all of the markers which are associated with pluripotent stem cells in the human fetal ovary (Kerr et al., 2008). Specific subpopulations of PGCs and oogonia expressing OCT4, NANOG, and c-KIT markers were identified and stage-specific embryonic antigen-4 (SSEA-4) expression was found throughout the entire human fetal ovary by immunohistochemistry.
Commitment of PGCs to germ cells during mammalian embryogenesis

Molecular mechanisms which enable the commitment of PGCs to germ cells during mammalian embryogenesis are still poorly understood due to the limited amount of available fetal ovarian tissue to perform complex research. Only a few genes (TNAP, BLIMP1, STELLA, and FRAGILIS) are known to mark the germ cell commitment in the outer layer of cells of the embryo epiblast and can be used with some success to detect PGC formation in vitro. Recently, 11 genes specifically expressed in female and male mouse fetal germ cells, but not in ESCs and somatic tissues, were identified (Sabour et al., 2011). These genes were: Fkbp6, Mov1011, 4930432K21Rik, Tex13, Akt3, Gm1673, Hba-a1, Pik3r3, Ploc2, Spo11, and Tdrkh. Three of these genes - 4930432K21Rik, Tex13 (testis-expressed gene 13) and Gm1673 (predicted gene 1673) are novel and their functions are still unknown. These genes have already been proposed as ideal and specific markers to identify germ cells developed in vitro from pluripotent stem cells (Sabour et al., 2011). FIGLA and NOBOX genes represent a growing number of oocyte-specific transcription factors which regulate different genes unique to early oogenesis (Pangas and Rajkovic, 2006).

Stemness of PGCs

Because of their stem-like character, human PGCs and EGCS have already been proposed to be researched in terms of cell therapies and potential development into oocytes (Aflatoonian and Moore, 2005). Mature oocytes can be generated in vitro from the mouse oogoniaia (Qing et al., 2008), human primary oocytes obtained by mechanical disaggregation and cultured in a SCF medium (Brieno-Enriquez et al., 2010), bovine primordial follicles (McLaughlin and Telfer, 2010), and also from mouse premeiotic fetal germ cells. A simple and efficient method as a combination of in vivo transplantation and in vitro culture (maturation) that can be used to obtain mature oocytes from the premeiotic germ cells of a fetal mouse has been developed (Shen et al., 2006). By using this method, mouse fetal ovaries were isolated and transplanted under the kidney capsule of the recipient mice to initiate oocyte growth from the premeiotic germ cells, and they were recovered 14 days later. Subsequently, the primary and early secondary follicles generated in the ovarian grafts were isolated and cultured in vitro for 16 days. The mature oocytes “ovulated” from these follicles and were able to be fertilized in vitro and to produce live offspring. The offspring after the in vitro fertilization were normal and were able to successfully mate with both females and males. The patterns of the methylated sites of the in vitro matured oocytes were similar to those of normal mice. It has been shown that the whole process of oogenesis, from premeiotic germ cells to germinal vesicle-stage oocytes, can be carried out under the kidney capsule of the recipient mice (Shen et al., 2006). Moreover, recently a population of PGCs was collected from the mouse female fetal gonads and, together with gonadal somatic cells, transplanted under the kidney capsule of adult mice (Matoba and Ogura, 2011). The transplanted cells formed ovarian-like tissue under the kidney capsule, and fully grown germline vesicle oocytes developed within this tissue. PGC-derived oocytes were isolated, matured in vitro, and microinjected with normal sperm. In this way the retrieved oocytes were fertilized, and after the transfer of the embryos into the uterus of adult mice, normal pups were born (Matoba and Ogura, 2011). This work demonstrated the flexibility of the PGCs' development into the competent female gametes and proposed the transplantation procedure as a technical basis for the induction of the development of early germ cells of exogenous origins, such as those from embryonic stem cells. Mouse fetal germ cells were also capable of forming primordial follicles and developing into mature oocytes in vitro, which can be successfully fertilized in vitro and developed into embryos at the morula/blastocyst stage (Shen et al., 2007).

All this methodology could not be used in human reproductive medicine, but brought important basic knowledge which could support the successful in vitro differentiation of pluripotent stem cells retrieved from different sources into the competent oocytes in future.

Stem cells in adult mouse ovaries

The existence of female germline stem cells (GSCs) in postnatal mammalian ovaries still remains a dilemma and a controversial issue among reproductive biologists and stem cell researchers (Skaznik-Wiikiel et al., 2007; Tilly and Telfer, 2009). After some previous indirect evidence about the presence of stem cells in adult mouse ovaries (Johnson et al., 2004, 2005) an important breakthrough was made by Zou and his co-workers (Zou et al., 2009). They found the presence of Mvh (mouse Vasa homologue) positive cells in the ovarian surface epithelium of neonatal mice ovaries. After immunomagnetic isolation, they established a neonatal mouse germ stem cell (mGSC) line persisting for more than 15 months. At the same time, GSCs from adult mouse ovaries were isolated and cultured for more than 6 months. These GSCs retained high telomerase activity and a normal karyotype during prolonged culture. After infection with a green fluorescent protein (GFP) virus and its transplantation into the infertile mice, transplanted GSCs underwent oogenesis and the mice produced GFP transgene marked offspring (Zou et al., 2009).

By using a transgenic mouse model in which the GFP was expressed under a germ cell-specific Oct4 promoter, multipotent stem cell lines were isolated and established from the mouse postnatal and adult ovaries (Pacchiarotti et al., 2010). Two distinct populations of
GFP-\textit{Oct4} positive cells were found in the mouse ovaries, based on their distribution and size. A small group of cells with an average diameter of 10-15 \textmu m was located at the ovarian surface epithelium and larger cells with an average diameter of 50-60 \textmu m resembling oocytes were located in the center of the follicular compartment. Flow cytometry analysis revealed that the percentage of GFP-\textit{Oct4} positive cells in the mouse ovaries significantly decreased with age; while 1-2\% positive cells were found in the neonatal mice ovaries, only 0.05\% was still present in the adult ovaries (Pacchiarotti et al., 2010). These ovarian GSC lines maintained their stem cell characteristics, high telomerase activity, and normal karyotype after many passages for more than 1 year. They formed embryoid body-like structures with a differentiation into all three germ cell layers (endoderm, mesoderm, and ectoderm). The germline stem cells were distinct from the CD133-positive cells circulating in the bloodstream.

Recently, two lines of colony-forming cells isolated from adult mouse ovaries were established on somatic fibroblasts, which expressed markers specific for pluripotent embryonic stem cells and formed embryoid bodies and teratoma after injection into SCID mice (Gong et al., 2010). The embryonic stem cell-like cells in adult ovaries were proposed to be researched also in humans, because they may represent an alternative for establishing autologous stem cell lines from adults without any genetic manipulation.

In all the above-mentioned studies, mouse ovarian stem cells were cultured in comparable culture conditions: in DMEM (Dulbecco’s Modified Eagle’s Medium) or MEM-\textalpha (Minimal Essential Medium-alpha) culture medium containing fetal bovine serum (FBS), antibiotics (penicillin, streptomycin) and some other substances (i.e., sodium pyruvate, non-essential amino acids, L-glutamine, \beta-mercaptoethanol, LIF, transferrin, insulin, putrescine, EGF, GDNF, basic FGF), and on mitotically inactivated mouse embryonic fibroblasts (MEFs), as can be seen in Table 1.

The characterization, culture, and in \textit{vivo} and in \textit{vitro} differentiation of putative thecal stem cells isolated from the mouse neonatal ovaries have already been published (Honda et al., 2007).

There is also some evidence that aged mouse ovaries possess not only stem cells, but also dormant premeiotic germ cells expressing \textit{Stra8} and Dazl genes that can develop into fully competent GFP-positive oocytes following transplantation into a young host mouse environment, as revealed by the use of aged germline-specific GFP-expressing transgenic mice (Niikura et al., 2009).

**Stem cells in adult human ovaries**

Very little is known about the presence of stem cells in adult human ovaries. One of the main reasons is that human ovarian tissue is not an easily available material to be researched for the presence of stem cells. In spite of this, Bukovsky and his co-workers scraped the ovarian surface epithelium of postmenopausal ovaries and cultured the scraped population of cells \textit{in vitro} (Bukovsky et al., 2005). Although human ovarian surface epithelium cell cultures have been previously cultured in terms of research of its biology and epithelial cancer formation, Bukovsky’s group was the first to observe the development of large oocyte-like cells in postmenopausal ovarian surface epithelium cell cultures in the presence of estrogentic stimuli (phenol red). These large cells with a diameter of 180 \mu m exhibited germinal vesicle breakdown, expulsion of the polar body, and a surface expression of zona pellucida proteins, as revealed by cytochemistry. This was the indirect evidence of putative stem cells in the ovarian surface epithelium layer of postmenopausal ovaries with no naturally present follicles/oocytes. In the next step, they found the steroid-mediated differentiation of neural/neuronal cells from the epithelial ovarian precursors \textit{in vitro} (Bukovsky et al., 2008).

Some further steps were made by Virant-Klun and her co-workers. They identified an unknown population of putative stem cells in the ovarian sections \textit{in situ} (Fig. 1) and in a population of cells scraped from the ovarian surface epithelium (Fig. 2) of women with no naturally present follicles/oocytes - postmenopausal women and women with premature ovarian failure (Virant-Klun et al., 2008). These cells expressed some transcription factors of pluripotent embryonic stem cells such as OCT4, SOX2 and NANOG, and developed \textit{in vitro} into oocyte-like cells which expressed some oocyte-specific markers. By magnetic-activated cell sorting (MACS), two different types of SSEA-4 positive cells (Figure 2) were isolated by anti-SSEA-4 antibody-coated beads from the scraped ovarian surface epithelium in these women. These cells were positive for SSEA-4 surface antigen. Moreover, they found parthenogenetic blastocyst-like structures (Fig. 3) in the ovarian surface epithelium cell cultures of postmenopausal women with no naturally present follicles/oocytes (Virant-Klun et al., 2009; Virant-Klun and Skutella, 2010).

Furthermore, the multipotent subpopulation of luteinizing granulosa cells was isolated from the follicles, the follicular fluid of infertile women, included in the \textit{in vitro} fertilization programme (Kossowska-Tomaszczuk et al., 2009). These cells were maintained in culture over prolonged periods of time in the presence of leukemia-inhibitory factor (LIF), expressed mesenchymal lineage markers (CD29, CD44, CD90, CD105, CD117, and CD166) and were differentiated \textit{in vitro} into different cell types, such as neurons, chondrocytes, and osteoblasts. After their transplantation into immunodeficient (SCID) mice, these cells survived and generated \textit{in vivo} tissues of mesenchymal origin. There is also some new indirect evidence about the potential stem cells in the ovary in terms of telomerase activity in adult human ovaries (Liu and Li, 2010).
In vitro oogenesis

Enormous effort has been put into the development of oocytes from pluripotent embryonic stem cells (ESCs), which have an unlimited self-renewal feature and are able to differentiate into almost every mature cell type in the body (Öktem and Oktay, 2008). Some studies showed that blastocyst inner cell mass (ICM) cells expressed NANOS1, STELLAR and OCT4 genes, whereas undifferentiated human ESCs expressed all these genes along with the germ cell-specific gene DAZL (Clark et al., 2004). It is known that ESCs express some germ cell specific genes such as OCT4, BLIMP1, STELLA, FRAGILIS, VASA and DAZL. Upon ESC differentiation into embryoid bodies they expressed some RNA and protein markers of immature premeiotic germ cells and mature germ cells, including VASA, BOL, SCP1, SCP3, GDF9, and TEKT1 (Clark et al., 2004).

ESCs can develop into oocyte-like cells given the right conditions, but most of the studies confirmed that these cells were not mature and fully competent oocytes. Oocytes developed in vitro expressed only some oocyte-specific genes, did not extrude zona pellucida and did not progress through the process of meiosis in most cases.

In 2003 it was published in the Science journal that mouse embryonic stem cells (mESCs) had been for the first time successfully developed into oocytes (Hübner et al., 2003). These cells expressed some oocyte specific genes and seemed to enter meiosis; they recruited adjacent cells to form follicle-like structures, and were able to develop into parthenogenetic blastocysts. Therefore, an advanced study of this possible approach to infertility treatment was proposed (Kehler et al., 2005). Later, it was reported that mESCs indeed formed follicle-like ovarian structures with oocyte-like cells.

<table>
<thead>
<tr>
<th>Source</th>
<th>Isolation</th>
<th>Culture medium and feeder layer</th>
<th>Culture conditions</th>
<th>Cell morphology and function</th>
<th>Cell characteristics</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>5 day-old (neonatal) and adult mice ovaries</td>
<td>-two-step enzymatic digestion -MACS isolation of Mvh (mouse Vasa homologue) positive cells</td>
<td>MEM-κ containing FBS, sodium pyruvate, non-essential amino acids, L-glutamine, β-mercaptoethanol, LIF, transferrin, insulin, putrescine, mouse EGF, human GDNF, human basic FGF, penicillin</td>
<td>-medium changed every 2-3 days</td>
<td>-large round or ovoid cells, nuclear diameter of 12-20 µm, little cytoplasm</td>
<td>-expressed markers: Oct4, Mvh, Dazl, Blimp1, Fragilis, Stella, Rex1</td>
<td>Zou et al., 2009</td>
</tr>
<tr>
<td>F1 (C57BL/6XCD-1) hybrid mice</td>
<td>Mice were injected with BrdU for cell proliferation analysis.</td>
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<tr>
<td>Adult (10-week old) mice ovaries</td>
<td>-digestion by trypsin-EDTA and collagenase Type I -filtration and centrifugation</td>
<td>DMEM containing FBS, non-essential amino acids, L-glutamine, β-mercaptoethanol, mouse LIF, lyophilized mixture of penicillin and streptomycin MEF (mitotically inactivated mouse embryonic fibroblasts)</td>
<td>-on day 7 of primary culture cell colonies mechanically removed and placed on a new MEF monolayer -medium changed daily -sub-passaged every 3 days</td>
<td>-colony-forming cells in ovarian stroma</td>
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<tr>
<td>F1 (C57BL6XD2A) hybrid mice and outbred (ICR) mice</td>
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<td>Differentiated in vitro into neuronal cells.</td>
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<td>Neonatal GSCs (neonatal) and adult ovaries</td>
<td>-digestion by collagenase and DNase-I -GFP fluorescent-activated cell sorting (FACS)</td>
<td>According to Okamoto et al., 1990: MEM-κ containing FBS MEF (mitotically inactivated mouse embryonic fibroblasts)</td>
<td>-on day 7, small colonies manually split for 4-5 passages before using trypsin -half of medium changed every other day</td>
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<td>OG2 mice</td>
<td>Every two weeks cells transferred to a new MEF plate.</td>
<td></td>
<td></td>
<td>Two distinct populations of cells with different diameters: 1.) 10-15 µm in the ovarian surface epithelium and 2.) 50-60 µm in the center of the follicular compartment.</td>
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<td></td>
<td></td>
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<td></td>
<td>Formed EB in vitro and did not form teratoma in SCID mice.</td>
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<tr>
<td>2 to 5 day-old (neonatal) and adult mouse ovaries</td>
<td>-digestion by collagenase and DNase-I -GFP fluorescent-activated cell sorting (FACS)</td>
<td>According to Okamoto et al., 1990: MEM-κ containing FBS MEF (mitotically inactivated mouse embryonic fibroblasts)</td>
<td>-on day 7, small colonies manually split for 4-5 passages before using trypsin -half of medium changed every other day</td>
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which did not progress through the process of meiosis (Novak et al., 2006). It was found that whereas some meiotic genes like \textit{Scp3} were expressed in oocyte-like cells, some other meiotic proteins, such as \textit{Scp1}, \textit{Scp2}, \textit{Stag3}, \textit{Rec8} and \textit{Smc1} were not expressed. It was concluded that ESC-derived oocyte-like cells showed the absence of essential meiotic proteins and failed to progress through meiosis. Oocyte-like cells and follicle-like structures expressing oocyte-specific genes such as \textit{Figa} (\textit{Figa} and \textit{Zp3}) have been developed from the mESCs cultured in conditioned medium collected from newborn male testicular cell cultures (Lacham-Kaplan et al., 2013).
Fig. 2. Putative stem cells (arrows) in the human ovarian surface epithelium scrapings of patients with premature ovarian failure. A-C. Putative stem cells (arrows) among clusters of scraped epithelial cells or attached to epithelial cells (inverted microscope, Nomarski, immersion objective, magnification x 6000). D. Putative stem cells (arrows) among cluster of scraped epithelial cells (inverted microscope, Hoffman, magnification x 200). E-H. Two different populations of SSEA4-positive cells isolated from the ovarian surface epithelium scrapings by anti-SSEA-4 antibody-coated beads and magnet: population of bigger cells (E, F) with diameters ~10 µm and smaller cells (G, H) with diameters from 2 to 5 µm (inverted microscope, Nomarski, immersion objective, magnification x 1000). I. SSEA-4 positive cells isolated by anti-SSEA-4 antibody-coated beads and magnet from the ovarian surface epithelium scrapings (fluorescent microscope, magnification x 200) (Scale bars: 5 µm).
Fig. 3. Oocyte-like cells and blastocyst-like structures developed in the human ovarian surface epithelium cell cultures in postmenopausal women according to Virant-Klun et al., 2009. 

A. Growing oocyte-like cell with attached putative stem cell (arrow) (inverted microscope, Hoffman, magnification x 200).

B. Oocyte-like cell among other cells detached from the dish bottom by trypsin-EDTA (inverted microscope, Hoffman, magnification x 400).

C. Oocyte-like cell with a diameter of 95 µm (inverted microscope, Hoffman, magnification x 400).

D. Accumulation of tubulin resembling the meiotic spindle (arrow) under the surface membrane of the oocyte-like cell (confocal microscope, magnification x 400).

E. Blastocyst-like structure with inner cell mass-, trophectoderm-, and blastocoel cavity-like structures (inverted microscope, Hoffman, magnification 200x).

al., 2006). It was suggested that this methodology should be applied to other mammalian species, including humans.

Not only ESCs isolated from embryos can develop into oocyte-like cells at the appropriate condition. After more publications (Dyce and Li, 2006a,b; Linher et al., 2009) indicating that primordial germ-like cells and oocyte-like cells differentiated in vitro from porcine fetal skin-derived stem cells, Dyce and co-workers recently reported the results of a more complex analysis of oocyte-like cells differentiated in this way (Dyce et al., 2010). They found that oocyte-like cells derived from the fetal porcine skin stem cells expressed some oocyte-specific genes like Oct4, Dazl, Vasa, Gdf9b, genes ZpB and ZpC for zona pellucida, and meiotic genes Dmc1 and Scp3, whereas they did not express Rec8 and Stra8 meiotic genes. They concluded that skin-derived stem cells from both female and male porcine fetuses were capable of entering into the oocyte differentiation pathway, but their current culture system was inadequate to support the complete development of competent oocytes. Moreover, Danner and co-workers derived oocyte-like cells expressing some germ cell genes (i.e., Vasa, Gdf9, SCP3, Bnc1) from rat clonal pancreatic stem cell lines (Danner et al., 2007).

Recently, it was confirmed that induced pluripotent stem cells (iPSCs) might also be developed into oocyte-like cells in vitro. Mouse iPSC cells derived from adult hepatocytes were able to differentiate into oocyte-like cells marked by mouse Vasa homolog Mvh expression in

### Table 2. Differentiation of embryonic stem cells into oocyte-like cells according to the literature.

<table>
<thead>
<tr>
<th>Source (XX)</th>
<th>Stem cell culture medium</th>
<th>Feeder layer</th>
<th>Oocyte-like cell maturation medium</th>
<th>Transfection/other manipulation</th>
<th>Expressed oocyte-specific markers</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>mESCs</td>
<td>DMEM containing glucose, FCS, L-glutamine, non-essential amino acids, β-mercaptoethanol, penicillin, streptomycin, murine LIF</td>
<td>MEFs (mitotically inactivated mouse embryonic fibroblasts) in 0.1% gelatine-coated plates</td>
<td>MEM-α supplemented with BSA, pyruvic acid, transferrin, selenium, insulin, EGF, gonadotropins, hGC and PMSF</td>
<td>Yes. mESCs were transfected with gcOct4-GFP.</td>
<td>Oct4, Vasa, c-kit, Dmc1, Scp3, Gdf9, Zp1, Zp2, Zp3, Figla</td>
<td>Hübner et al., 2003</td>
</tr>
<tr>
<td>mESCs line R1, passage 14</td>
<td>DMEM containing glucose, FBS, L-glutamine, non-essential amino acids, β-mercaptoethanol, penicillin, streptomycin, LIF</td>
<td>MEFs (mitotically inactivated mouse embryonic fibroblasts) in 0.1% gelatine-coated tissue culture plates</td>
<td>Supplemented DMEM medium without LIF and MEFs</td>
<td>Yes. Cells cocultured with a stable mouse 3T3 fibroblast cell line overexpressing BMP4 and transfection to generate a stable mouse BMP4 cell line.</td>
<td>SSEA-1, Scp3, Zp3</td>
<td>Novak et al., 2008</td>
</tr>
<tr>
<td>mESCs</td>
<td>DMEM containing FCS, non-essential amino acids, β-mercaptoethanol, penicillin, streptomycin, mouse recombinant LIF</td>
<td>0.1% gelatine coated plastic flakes</td>
<td>Testicular Cell Conditioned (TCC) medium prepared from testicular tissue of 1-day-old newborn male mice testes</td>
<td>No.</td>
<td>Oct34, Mvh, c-kit (very few), Stella, Dazl, Figla, Zp3, -did not express Zp1 and Zp2 -formed embryoid body-like structures</td>
<td>Lacham-Kaplan et al., 2006</td>
</tr>
<tr>
<td>hESCs</td>
<td>HES-3 (XX) Basic medium: 90% DMEM containing FCS, L-glutamine, antimalocytics</td>
<td>POF (mitotically inactivated porcine ovarian fibroblasts)</td>
<td>POF-conditioned medium with/without forskolin and retinoic acid</td>
<td>No.</td>
<td>VASA, BOULE, DAZL, GDF3, GDF9, MLH1, PUM2, OCT4 -did not express SCP1 -formed EB-like structures, multilineage differentiation (AFP, NESTIN)</td>
<td>Richards et al., 2010</td>
</tr>
<tr>
<td>hOSCs</td>
<td>DMEM/F-12 with phenol red, NaHCO3, FBS, penicillin, streptomycin, gentamycin</td>
<td>Native human ovarian fibroblasts from OSE scrapings</td>
<td>DMEM with phenol red containing NaHCO3, penicillin, streptomycin</td>
<td>No.</td>
<td>ZP, CK5, 6, 8, 17, CK18, VIMENTIN, PS1</td>
<td>Bukovsky et al., 2005</td>
</tr>
<tr>
<td>hOSCs</td>
<td>Basic medium: DMEM/F-12 with phenol red, NaHCO3, penicillin, streptomycin, gentamycin</td>
<td>Native human ovarian fibroblasts from OSE scrapings</td>
<td>Basic medium containing human follicular fluid</td>
<td>No.</td>
<td>SSEA-4, c-KIT, OCT4, SOX2, NANO2, c-KIT, ZP2, VASA -SCP3 was not expressed -formed blastocyst-like structures with normal chromosomes X,Y,13,16,18,21,22</td>
<td>Virant-Klun et al., 2008, 2009</td>
</tr>
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</table>

hOSCs: human ovarian stem cells from adult ovarian surface epithelium (OSE) scrapings, FCS: fetal calf serum, EB: embryoid body.
feeder-free and suspension cultures (Imamura et al., 2010). Moreover, an international group of experts tried to find the potential of human fetal and adult iPSCs derived from somatic cells to form primordial and meiotic germ cells, relative to hESCs (Panula et al., 2011). They found that approximately 5% of human iPSCs differentiated to PGCs following induction with BMPs. These PGCs expressed GFP from a germ cell-specific reporter and expressed endogenous germ cell specific proteins and mRNAs. In response to the overexpression of the intrinsic regulators, iPSCs formed meiotic cells with extensive synaptonemal complexes (SC) and post-meiotic haploid cells with a similar pattern of acrosin staining as observed in human spermatids. The female germ cells still need to be elucidated.

Factors affecting in vitro oogenesis

According to published reports, there are some factors, such as the appropriate selection of the feeder layer, the addition of the granulosa cells and other cells which can secrete BMP4, and the addition of retinoic acid (RA) into the culture medium, which could to some extent induce the differentiation of ESCs into the germ cells (Zhou et al., 2010). KIT ligand and BMP protein signalling enhances differentiation of hESCs into germ-like cells (West et al., 2010). The loss of KIT ligand in differentiation of cell cultures results in intense down-regulation of germ cell specific genes and a 70.5% decrease in germ-like cells expressing DDX4 and OCT4 (POUSF1) genes. On the other hand, endogenous BMP signalling caused germ-like cell differentiation. The inhibition of this pathway causes a significant decrease in the number of germ cell-like cells and in the expressions of germ cell specific genes (West et al., 2010). By eliminating feeders but maintaining their secreted extracellular matrix it is possible to sustain the increased numbers of germ cell-like cells in culture (West et al., 2010).

In most studies, oocyte-like cells were cultured in DMEM culture medium with different supplements (FCS or FBS, glucose, L-glutamine, non-essential amino acids, β-mercaptoethanol, LIF, NaHCO3, antibiotics, antimycotics) on mitotically inactivated mouse embryonic fibroblasts (Table 2). Only a few of them used some ovarian components in their culture systems, like mitotically inactivated porcine ovarian fibroblasts, natural human ovarian fibroblasts, follicular fluid and granulosa cells.

A comparative evaluation of different in vitro systems that stimulate germ cell differentiation in hESCs has already been performed (Richards et al., 2010). Embryoid bodies derived from hESC lines were cultured in six different culture conditions: mitotically inactivated porcine ovarian fibroblasts, a 100% conditioned medium from mitotically inactivated porcine ovarian fibroblasts, a 50% conditioned medium from mitotically inactivated porcine ovarian fibroblasts, forskolin, transretinoic acid, and forskolin combined with retinoic acid. Mitotically inactivated porcine ovarian fibroblasts proved to be the best culture system to induce the hESCs differentiation into the germ cell direction by increasing the expression of several germ cell specific genes in embryoid bodies. Some researchers were able to isolate putative GSCs from embryoid bodies produced by ESCs by using Percoll and Nycodeinz density gradient centrifugation (Saiti and Lacham-Kaplan, 2008). In the mouse model, the positive effect of the conditioned medium of mesenchymal stem cells on the in vitro maturation of oocytes was confirmed (Ling et al., 2008). In addition to the self-renewal and multiple differentiations, mesenchymal stem cells (MSCs) secrete a variety of growth factors and cytokines beneficial for oocyte in vitro maturation. A large-scale production of growing oocytes from neonatal mouse ovaries was developed in vitro in a follicle-free culture system and at the sequential provision of essential nutrients and growth factors (Honda et al., 2009). At this point, about 800 otherwise dormant oocytes from a newborn mouse developed, reached the size of oocytes in normal antral follicles, entered the metaphase I meiotic stage, extruded zona pellucida and were capable of fusing with sperm.

It has already been demonstrated that female germ-like cells can be derived from the ESCs through the formation of embryoid bodies. Yu et al. (2009) reported on the transgene expression approach to derive female germ cells directly from the mESCs in vitro without the formation of embryoid bodies. They confirmed the development of female germ cells through the ectopic expression of Dazl, which is an important germ-cell specific RNA-binding protein. They proposed Dazl as a master gene controlling germ cell differentiation.

Another study confirmed that mouse granulosa cells were effective in inducing the differentiation of mESC-derived PGCs into oocyte-like cells through direct cell-to-cell contacts (Qing et al., 2007).

Oocyte maturation from ESCs by transplantation

Some researchers preferred transplantation, which directs oocyte maturation from the ESCs and could provide a new strategy for female infertility treatment in the future.

Progress has been made by Reijo Pera’s group (Nicholas et al., 2009). For the first time, they differentiated mESCs, derived from transgenic mice carrying Oct4-GFP, into oocyte-like cells in the presence of a germ cell maturation factor cocktail (FAC) comprising antiapoptotic, germ cell specification and meiotic induction factors, including BMP4 protein, retinoic acid, cytochrome p450, CYP26 inhibitor, stromal cell-derived factor 1 (SDF1), stem cell factor, basicFGF, n-acetyl-cysteine and forskolin. ESC-derived oocytes expressed several oocyte-specific genes as revealed by single-cell gene expression analyses. Genetic analyses have also shown the requirement of Dazl expression for in vitro oocyte development and
limited in vitro maturation of ESC-derived oocytes. They observed that approximately 1-3% of Oct4-GFP positive cells initiated meiosis, as evidenced by synaptonemal complex protein (SCP) expression and chromosomal localization. However, they detected only partial chromosomal alignment of Scp3 and focal nuclear localization of Scp1 proteins in these oocytes as indicative of a limited meiotic progression. The most significant enhancement of meiosis induction in vitro was found when mESCs were differentiated in a coculture with dissociated mouse embryos although in vitro development of follicles was not observed. Because of the incomplete meiotic progression they tried to achieve ESC-derived oocyte maturation by transplantation into a synchronized mouse ovarian niche (Nicholas et al., 2009). To enable the ovarian niche, Oct4-GFP oocyte-like cells derived from mESCs were co-aggregated with cells retrieved by enzymatic dissociation of mouse neonatal ovaries and transplanted under the kidney capsule of bi-laterally ovariectomized SCID recipient mice. The formation of primary follicles with oocytes was observed. They concluded that considerable work still remains to be done before safe and effective clinical translation could be realized (Nicholas et al., 2009).

Even in vitro maturation of immature - germinal vesicle oocytes retrieved in the in vitro fertilization programme is not a sufficiently efficient procedure. ESC-derived oocyte maturation definitely fails in the condition in vitro, therefore the transplantation of these oocyte-like cells into an ovarian niche to naturally direct their function and maturity or to 'wake up' the ovaries seems to be the best option to achieve the fully competent oocyte, and a potential therapeutic strategy for ovarian infertility.

Oocyte generation in adult mouse ovaries by putative germ cells in bone marrow and peripheral blood has also been evidenced (Johnson et al., 2005; Lee et al., 2007). After bone-marrow transplantation via the tail vein, cell tracking showed that donor mice-derived oocytes were generated in ovaries of recipient mice with ovarian failure due to cytostatic busulfan treatment. In human medicine, there have been some clinical reports about fertility recovery and pregnancy after bone marrow transplantation in patients with ovarian failure due to high-dose chemotherapy (cyclophosphamide, busulfan) or total-body irradiation because of hematologic malignancy (Sanders et al., 1996). Similarly, ovarian recovery and pregnancy have been documented after hematopoietic stem cell transplantation in some Fanconi anemia patients who were infertile due to ovarian damage from myeloablative conditioning (Nabhan et al., 2010). The mechanisms of ovarian recovery in these patients remain unknown.

**Ovarian stem cells and cancer**

Pluripotent/multipotent stem cells present in adult human ovaries might be somehow involved in the female reproductive function. With inappropriate conditions in the body, they might induce the formation of tumors and the development of cancer in humans. Also, in the most aggressive epithelial ovarian cancers a subpopulation of stem-cell like cells was found which expressed transcription factors LIN28 and OCT4 characteristic for pluripotent stem cells (Peng et al., 2010). Both epithelial ovarian cancer cell lines and the patient’s tumor samples expressed these transcription factors which are usually highly expressed in hESCs. The combined expression of these two proteins in tumor samples was correlated with an advanced tumor grade. When these two proteins were repressed in the same cells by using RNA interference there was a significant reduction of cancer cell growth and survival (Peng et al., 2010). Therefore, it was proposed that these factors of pluripotency may serve as important molecular diagnostics and therapeutic targets for the development of new treatment approaches in patients with aggressive ovarian epithelial cancers. Similarly, Ye et al. (2010) found the expression of another pluripotent stem cell marker, SSEA-4 in solid tumors of patients with epithelial ovarian carcinomas. It was proposed that SSEA-4 may play a role during the oncogenesis of epithelial ovarian carcinoma and may be a potential therapy target in these patients. Additionally, OCT4 expression can be found in immature neuroepithelium and reflect the differentiation of neural tissue in immature teratoma of the ovary (Abiko et al., 2010).

**Conclusion**

Differentiation of oocytes from the pluripotent ESCs/iPSCs has the potential to becoming a future source of oocytes for research use (Marques-Mari et al., 2009) to better understand the molecular mechanisms of oogenesis, oocyte maturation, fertilization, and embryo development. In this way, the pathologies resulting in human infertility would be better elucidated. Additionally, if all complex genetic and epigenetic methodological limitations could be safely solved and complex culture conditions and an appropriate niche could be established, some therapeutic opportunities could be considered. The development of autologous ESCs/GSCs into mature oocytes in vitro would enable in vitro fertilization and a desired baby with patients with severe ovarian infertility and no naturally present follicles/oocytes (i.e. premature ovarian failure) in the future. It has also been proposed that pluripotent/multipotent stem cells may play a role during the oncogenesis of epithelial ovarian cancers and may be a potential therapy target in these patients to provide more efficient therapy in the future.

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